





# Allergen immunotherapy for allergic rhinoconjunctivitis: A systematic review and meta-analysis

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#### Abstract

**Background:** The European Academy of Allergy and Clinical Immunology (EAACI) is in the process of developing Guidelines on Allergen Immunotherapy (AIT) for Allergic Rhinoconjunctivitis. To inform the development of clinical recommendations, we undertook a systematic review to assess the effectiveness, cost-effectiveness, and safety of AIT in the management of allergic rhinoconjunctivitis.

**Methods:** We searched nine international biomedical databases for published, in-progress, and unpublished evidence. Studies were independently screened by two reviewers against predefined eligibility criteria and critically appraised using established instruments. Our primary outcomes of interest were symptom, medication, and combined symptom and medication scores. Secondary outcomes of interest included cost-effectiveness and safety. Data were descriptively summarized and then quantitatively synthesized using random-effects meta-analyses.

**Results:** We identified 5960 studies of which 160 studies satisfied our eligibility criteria. There was a substantial body of evidence demonstrating significant reductions in standardized mean differences (SMD) of symptom (SMD  $-0.53$ , 95% CI  $-0.63$ ,  $-0.42$ ), medication (SMD  $-0.37$ , 95% CI  $-0.49$ ,  $-0.26$ ), and combined symptom and medication (SMD  $-0.49$ , 95% CI  $-0.69$ ,  $-0.30$ ) scores while on treatment that were robust to prespecified sensitivity analyses. There was in comparison a more modest body of evidence on effectiveness post-discontinuation of AIT, suggesting a benefit in relation to symptom scores.

**Conclusions:** AIT is effective in improving symptom, medication, and combined symptom and medication scores in patients with allergic rhinoconjunctivitis while on treatment, and there is some evidence suggesting that these benefits are maintained in relation to symptom scores after discontinuation of therapy.

#### KEYWORDS

allergen, allergen immunotherapy, allergic rhinoconjunctivitis, subcutaneous, sublingual

## 1 | BACKGROUND

Allergic rhinoconjunctivitis is a very common chronic condition that can result in considerable morbidity and impairment of quality of life.<sup>1,2</sup> The disease is triggered by exposure to seasonal and/or perennial allergens and, depending on the nature of the allergenic trigger(s) and patterns of exposure, symptoms may be persistent or intermittent.<sup>3</sup> Allergic rhinitis is typically characterized by symptoms of nasal obstruction, a watery nasal discharge, sneezing and itching, and there is often (but not invariably) involvement of the conjunctiva (allergic conjunctivitis), which manifests with itching, injection and tearing.<sup>4</sup> There may in addition be an impact on the ability to concentrate, on school and work performance,<sup>5,6</sup> and interference with daily activities and sleep; furthermore, allergic rhinitis is a risk factor for the development of asthma.<sup>7</sup>

Symptoms can, in many cases, be controlled with avoidance measures and pharmacological therapies such as oral, intranasal and topical (ophthalmic) H<sub>1</sub>-antihistamines, intranasal corticosteroids and antileukotrienes, as monotherapy or in combination.<sup>8,9</sup> Allergen immunotherapy (AIT) is an additional potential treatment option, particularly for those with more troublesome disease which remains inadequately controlled despite avoidance measures and regular pharmacotherapy.<sup>8-10</sup> The problem of inadequately controlled allergic rhinoconjunctivitis, despite optimal medical treatment, continues to represent a therapeutic challenge in the majority of patients.

The European Academy of Allergy and Clinical Immunology (EAACI) is in the process of developing Guidelines on AIT for Allergic Rhinoconjunctivitis and this systematic review has been undertaken to inform the formulation of key clinical recommendations.

Specifically, we sought to assess the effectiveness, cost-effectiveness and safety of AIT in patients with allergic rhinoconjunctivitis.<sup>11</sup>

## 2 | METHODS

As our methods have been reported in detail in our published protocol,<sup>12</sup> we confine ourselves to a synopsis of the methods employed.

### 2.1 | Search strategy

A highly sensitive search strategy was developed and validated study design filters were applied to search nine electronic bibliographic databases. The search strategy was developed on OVID MEDLINE and then adapted for the other databases (see Appendix S1). In all cases, the databases were searched from inception to October 31, 2015. Additional references were located through searching the references cited by the identified studies, and unpublished work, while research in progress was identified through discussion with experts in the field. We invited experts from a range of disciplines and regions to add to the list of included studies by identifying additional published and unpublished papers they were aware of and research in progress. There were no language restrictions employed; where possible, relevant literature was translated into English.

### 2.2 | Inclusion criteria

We focused on studies conducted on patients of any age with allergic rhinoconjunctivitis investigating the effect of AIT. See Box 1 for full details.

#### Box 1 Inclusion and exclusion criteria

Patient characteristics	Studies conducted on patients of any age with a physician-confirmed diagnosis of allergic rhinoconjunctivitis or allergic rhinitis, plus evidence of clinically relevant allergic sensitization (eg, skin prick test or specific IgE).
Interventions of interest	Allergen immunotherapy (AIT) for different allergens (eg, pollen, house dust mites (HDM), animal dander, cockroach and molds), including modified allergens, administered through the subcutaneous immunotherapy (SCIT), sublingual immunotherapy (SLIT), intralymphatic immunotherapy (ILIT) or any other routes.
Comparator	Placebo or any active comparator.
Study designs	<i>Effectiveness:</i> Robust double-blind RCTs. Originally, we planned to include data from any RCT, irrespective of whether there was blinding. This was changed due to the volume of RCT studies. This decision was made prior to any analyses being undertaken. <i>Cost-effectiveness:</i> health economic analysis. <i>Safety:</i> Double-blind RCTs and large case series ( $\geq 300$ patients).
Study outcomes	<i>Primary outcomes:</i> effectiveness, both short-term (ie, during treatment) and long-term (ie, at least a year after discontinuation of AIT), as assessed by symptom and/or medication scores. <i>Secondary outcomes:</i> disease-specific quality of life (QoL); threshold of allergen exposure to trigger symptoms on allergen challenge or in an environmental exposure chamber; health economic analysis from the perspective of the health system/payer; and safety as assessed by local and systemic reactions in accordance with the World Allergy Organization's (WAO) grading system of side-effects. <sup>14,15</sup>
Exclusion criteria	Reviews, discussion papers, nonresearch letters and editorials, animal studies, and studies not employing double-blind RCT designs.

## 2.3 | Study selection

All references were uploaded into the systematic review software DistillerSR and underwent initial de-duplication. Study titles were independently checked by two reviewers (SD and UN) according to the above selection criteria and categorized as included, not included or unsure. For those papers in the unsure category, we retrieved the abstract and recategorized as above. Any discrepancies were resolved through discussion and, if necessary, a third reviewer (AS) was consulted. Full-text copies of potentially relevant studies were obtained and their eligibility for inclusion independently assessed by two reviewers (SD and UN). Studies that did not fulfill all of the inclusion criteria were excluded.

## 2.4 | Quality assessment strategy

Quality assessments were independently carried out on each study by two reviewers (UN, SA, AA, MA, or TM) using a range of instruments. RCTs were assessed for generation of allocation sequence, concealment of allocation, baseline outcome measurements, baseline characteristics, incomplete outcome data, blinding of outcome assessor, protection against contamination, selective outcome reporting and other risks of bias using the Cochrane Risk of Bias (ROB) Tool.<sup>13</sup> We used the Critical Appraisal Skills Programme (CASP) Economic Evaluation Checklist for health economic studies.<sup>14</sup> For case series, we used the quality assessment tool produced by the National Institute for Health and Clinical Excellence (NICE).<sup>15</sup> Any disagreements were resolved through discussion and, if necessary, a third reviewer (SD or AS) was consulted.

## 2.5 | Data extraction, analysis and synthesis

Data were independently extracted onto a customized data extraction sheet in DistillerSR by two reviewers (UN, SA, AA, MA, SD or TM), and any discrepancies were resolved by discussion or, if agreement could not be reached, by arbitration by a third reviewer (SD or AS). A descriptive summary with detailed data tables was initially produced to summarize the literature. Where clinically and statistically appropriate, meta-analyses were undertaken using random-effects modeling.<sup>16</sup> Data were extracted from primary studies, but where these were not available in a suitable format we first contacted authors for data and then if data were still not available we extracted data from previous Cochrane reviews. For outcomes for which it was not possible to produce a meta-analysis, we narratively synthesized data. Heterogeneity statistics are reported with each forest plot.

## 2.6 | Sensitivity analyses and assessment for publication bias

Sensitivity analyses were undertaken for the primary outcomes by comparing the summary estimates obtained by excluding studies considered to be at high ROB.

Publication bias was assessed for these same primary outcomes through the creation of funnel plots, and tested by Egger's regression test and Begg's rank correlation test.<sup>17,18</sup>

## 2.7 | Subgroup analyses

A number of subgroup analyses were undertaken, which are listed in the protocol.

## 2.8 | Registration and reporting

This review is registered with the International Prospective Register of Systematic Reviews (PROSPERO): <http://www.crd.york.ac.uk/prospero/>. The registration number is CRD42016035373. The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) checklist has been used to guide the reporting of this systematic review: <http://www.prisma-statement.org/> (Appendix 2, Supplementary file).

# 3 | RESULTS

Our search strategy yielded 5960 titles of which 160 studies (reported in 166 papers) met our overall review eligibility criteria. These eligible papers included 134 double-blind RCTs, 19 health economic analyses and seven case series (Figure 1).

## 3.1 | Effectiveness

### 3.1.1 | Description of trials

We identified 61 subcutaneous immunotherapy (SCIT) RCTs (reported in 63 papers)<sup>19-81</sup> including 6379 patients, 71 sublingual immunotherapy (SLIT) RCTs (reported in 75 papers)<sup>82-119,119-121,121-156</sup> including 13 636 patients and two intralymphatic immunotherapy (ILIT) RCTs<sup>157,158</sup> including 56 patients (Table 1a-c). The majority of studies only included adult participants. A range of allergens were assessed including weed, tree and grass pollens, molds, cat and dog dander, and house dust mites. A range of AIT protocols were utilized. The overwhelming majority of trials only reported on short-term effectiveness (Table S1a-c). A full description of the trials is given in the Data S1.

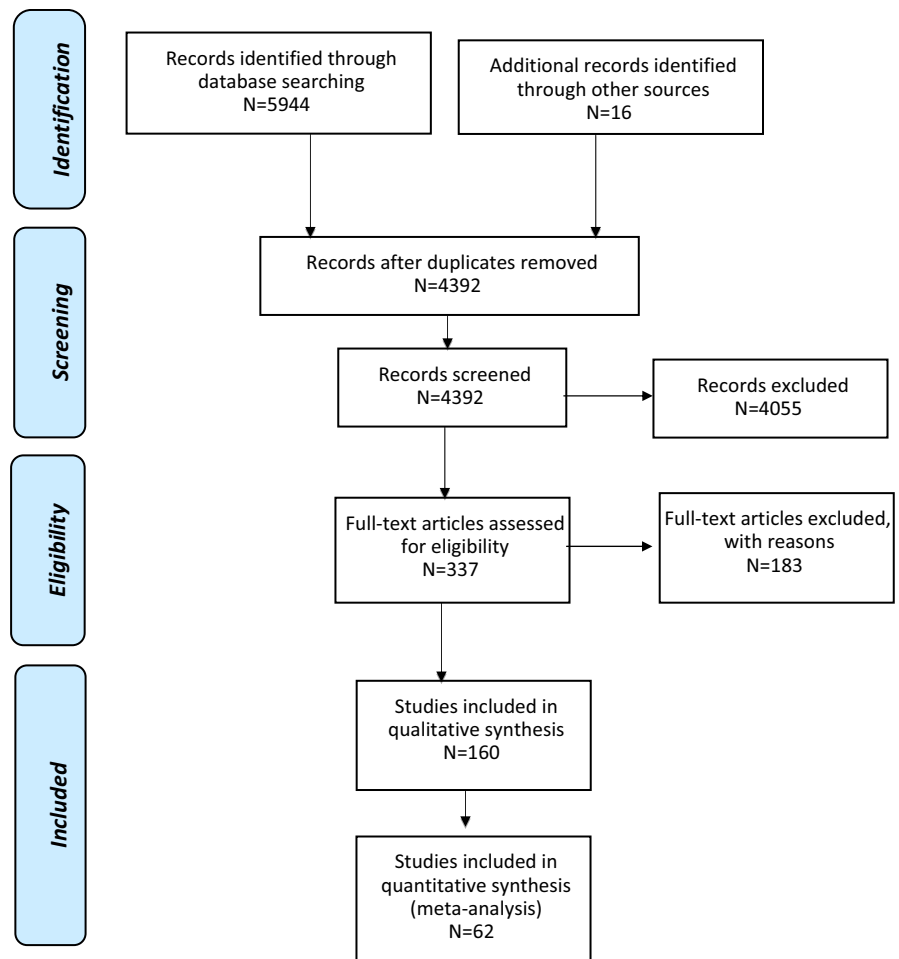
### 3.1.2 | Quality assessment

#### Subcutaneous immunotherapy

Overall, the quality of included studies was high. Thirty-seven studies were found to be at low ROB, eight studies at high ROB, and 16 were judged at unclear ROB (Table S1d).

#### Sublingual immunotherapy

The quality of studies was assessed to be low ROB in 26 studies, high ROB in 16 studies and unclear ROB in 28 studies (Table S1e). In one study, ROB could not reliably be assessed from the translation.



**FIGURE 1** PRISMA Diagram

### Intralymphatic immunotherapy

Both studies had a low ROB (Table S1f).

## 3.2 | Primary outcomes

Data on primary outcomes are summarized in Tables S1g-i.

### 3.2.1 | Symptom scores

#### Short-term

105 studies reported on the short-term effectiveness of AIT administered by the SCIT ( $n=51$ ), SLIT ( $n=52$ ) and ILIT ( $n=2$ ) routes assessed by symptom scores.

We were able to pool data from 58 SCIT and SLIT studies assessing the effectiveness of AIT by symptom scores. This showed a standardized mean difference (SMD) of  $-0.53$  (95% CI  $-0.63, -0.42$ ) this suggesting a moderate effect in favor of AIT (Figure 2).

**Sensitivity analysis** Sensitivity analysis was performed excluding all studies at high ROB, which demonstrated a SMD of  $-0.57$  (95% CI  $-0.68, -0.46$ ) (Figure S1).

**Assessment for publication bias** There was evidence of potential publication bias (Figure S2) which was also suggested by the Begg ( $P=0.003$ ) and Egger ( $P=0.003$ ) tests.

#### Subgroup analyses

Subgroup analyses were undertaken to compare:

- SCIT vs SLIT: SMD  $-0.65$  (95% CI  $-0.86, -0.43$ ) for SCIT and SMD  $-0.48$  (95% CI  $-0.61, -0.36$ ) for SLIT (Figures 3A and B), these both showing evidence of benefit; data from the two ILIT trials could not be pooled, but these studies also demonstrated an improvement in short-term symptom scores.
- Children vs adults for AIT (SCIT and SLIT): SMD  $-0.25$  (95% CI  $-0.46, -0.05$ ) for children and SMD  $-0.56$  (95% CI  $-0.70, -0.42$ ) for adults (Figures 4A and B), these analyses showing evidence of benefit in both adults and children.
- Children vs adults for SLIT only: SMD  $-0.42$  (95% CI  $-0.63, -0.21$ ) for children and SMD  $-0.47$  (95% CI  $-0.64, -0.29$ ) for adults (Figures S3A and B), these analyses showing benefit in both adults and children.
- Seasonal vs perennial allergens: SMD  $-0.37$  (95% CI  $-0.45, -0.28$ ) for seasonal and SMD  $-0.91$  (95% CI  $-1.47, -0.36$ ) for perennial (Figures S4A and B), these demonstrating evidence of benefit from both approaches.

**TABLE 1** (a) Characteristics of SCIT studies (n=61 studies, reported in 63 papers). (b) Characteristics of SLIT studies (n=71 studies, reported in 75 papers). (c) Characteristics of ILIT studies (n=2)

(a) Study (first author, Y, country)	Allergen no.										AIT Protocol				Product type/Name (manufacturer)				Short-term effectiveness		Long-term effectiveness		Quality of life									
	Grass pollen(s)	Tree pollen(s)	Weeds(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration		Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety		
Alvarez-Cuesta, 2005, Spain	X								X	X	X			X	X	X	X					1 yr	X	X	X	X	X	X	X	X	X	
Ariano, 1999, Italy		X						X	X	X				X	X	X	X					1 yr			X					X		
Arvidsson, 2002, Sweden	X							X	X	X				X	X	X	X	X				2 yr	X	X	X	X	X	X	X	X	X	
Balda, 1998, Germany	X							X	X	X				X	X	X	X					7 wk	X	X	X	X	X	X	X	X	X	
Bodtger, 2002, Denmark	X							X	X	X				X	X	X	X	X				1 yr	X	X	X	X	X	X	X	X	X	
Bousquet, 1987, France	X							X	X	X				X	X	X	X					10 mo	X	X	X	X	X	X	X	X	X	
Bousquet, 1989, France	X							X	X	X				X	X	X	X					8 mo	X	X	X	X	X	X	X	X	X	
Bousquet, 1990, France	X							X	X	X				X	X	X	X					NR	X	X	X	X	X	X	X	X	X	
Bousquet, 1991, France	X	X	X					X	X	X				X	X	X	X					1 yr	X	X	X	X	X	X	X	X	X	X
Bozek, 2016, Poland	X							X	X	X				X	X	X	X					3 yr	X	X	X	X	X	X	X	X	X	X
Brunet, 1992, Canada			X					X	X	X				X	X	X	X					3 mo	X	X	X	X	X	X	X	X	X	X
Ceuppens, 2009, Belgium & the Netherlands	X							X	X	X				X	X	X	X					18 mo	X	X	X	X	X	X	X	X	X	X

(Continues)

TABLE 1 (Continued)

(a) Study (first author, y, country)	Allergen(s) type					Allergen no.		Comparator					AIT Protocol				Product type/Name (manufacturer)			Short-term effectiveness		Long-term effectiveness			Quality of life								
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others)	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrasush	Rx duration	Symptom score	Medication score		Combined score	Symptom score	Medication score	Combined score	Safety			
Chakraborty, 2006, India	X								X	X	X	X	X	X	X	X	X					2 yr						X		X		X	
Charpin, 2007, France	X								X	X	X	X	X	X	X	X	X					15 mo						X		X		X	
Colas, 2006, Spain	X		X						X	X	X	X	X	X	X	X	X					1 yr						X		X		X	
Corrigan, 2005, UK	X								X	X	X	X	X	X	X	X	X					2 yr						X		X		X	
Crimi, 2004, Italy	X		X						X	X	X	X	X	X	X	X	X					3 yr						X		X		X	
Dokic, 2005, Macedonia		X							X	X	X	X	X	X	X	X	X					3 yr						X		X		X	
Dolz, 1996, Spain	X								X	X	X	X	X	X	X	X	X					3 yr						X		X		X	
Drachenberg, 2001, Germany and Austria	X								X	X	X	X	X	X	X	X	X					4-7 wk						X		X		X	
Drachenberg, 2002, Germany	X								X	X	X	X	X	X	X	X	X					4-7 wk						X		X		X	
DuBuske, 2011, USA, Canada, UK, Austria	X								X	X	X	X	X	X	X	X	X					4-8 wk						X		X		X	
Durham, 1999, UK Primary study Varney, 1991	X								X	X	X	X	X	X	X	X	X					3 yr						X		X		X	
Ewan, 1988, UK		X							X	X	X	X	X	X	X	X	X					3 mo						X		X		X	
Fell, 1988, UK	X								X	X	X	X	X	X	X	X	X					1 injection						X		X		X	

(Continues)



TABLE 1 (Continued)

(a) Study (first author, Y, country)	Allergen no.											Comparator			AIT Protocol				Short-term effectiveness		Long-term effectiveness										
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Product type/Name (manufacturer)	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety	Quality of life
Ferrer, 2005, Spain			X						X	X	X			X	X	X	X					20 mo	Biologically standardized extract of <i>Parietaria judaica</i> adsorbed onto aluminum hydroxide gel/Pangramin® Depot, ALK-Abelló	X	X	X			X	X	X
Frew, 2006, UK	X							X	X	X	X	X	X	X	X	X	X					1 yr	Standardized depot preparations of grass pollen extract/Alutard SQ grass pollen® (ALK-Abelló)	X	X	X			X	X	X
Grammer, 1982, USA			X					X	X	X	X	X	X	X	X	X	X					15 wk	Polymerized ragweed extract (PRW)/NR	X					X	X	X
Grammer, 1983, USA	X							X	X	X	X	X	X	X	X	X	X					4 mo	Six-grass pollen allergoid prepared by polymerization with glutaraldehyde/NR	X	X				X	X	X
Grammer, 1984, USA	X		X					X	X	X	X	X	X	X	X	X	X					>30 mo (UR)	Polymerized ragweed extract/NR			X			X	X	X
Grammer, 1987, USA	X							X	X	X	X	X	X	X	X	X	X					4 mo	Polymerized ragweed extract/NR			X			X	X	X
Höiby, 2010, Sweden & Germany	X							X	X	X	X	X	X	X	X	X	X					18 mo	Depigmented polymerized birch pollen (Betula alba) extract adsorbed onto aluminum hydroxide/Depigoid® (Laboratorios LETI S)			X			X	X	X
Horst, 1989, France				X				X	X	X	X	X	X	X	X	X	X			X		1 yr	Lyophilized and standardized Alt extract Stallergenes Laboratories			X			X	X	X
Iliopoulos, 1991, USA	X							X	X	X	X	X	X	X	X	X	X					~8 mo	Short ragweed extract/NR (Greer Laboratories, Lenoir, N.C.)			X			X	X	X
James, 2011, UK	X							X	X	X	X	X	X	X	X	X	X					2/4 yr	Phileum pratense extract adsorbed with aluminum hydroxide/Alutard SQ®			X			X	X	X
Juniper, 1990, Canada	X		X					X	X	X	X	X	X	X	X	X	X					6 wk	Modified ragweed tyrosine adsorbate/Pollinex® (Bencard Allergy Service)			X			X	X	X
Jutel, 2005, Poland	X							X	X	X	X	X	X	X	X	X	X			X		8-9 mo	Five recombinant grass pollen allergens/NR (Allergopharma)			X			X	X	X
Kleine-Tebbe, 2014, Spain, Germany & Austria	X							X	X	X	X	X	X	X	X	X	X					1 yr	Aluminum hydroxide-adsorbed Phileum pratense extract/AVANZ® Phileum pratense (ALK)	X	X	X			X	X	X
Klimek, 2014, Germany	X							X	X	X	X	X	X	X	X	X	X					1 yr	Glutaraldehyde-modified high polymerized allergen extract containing 6 grasses (60%) and rye pollen adsorbed onto aluminum hydroxide/CLUSTOID® (ROXALL Medizin)	X	X	X			X	X	X

(Continues)



TABLE 1 (Continued)

(a) Study (first author, y, country)	Allergen no.											AIT Protocol				Short-term effectiveness				Long-term effectiveness																						
	Allergen(s) type											Comparator				AIT Protocol				Short-term effectiveness				Long-term effectiveness																		
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Product type/Name (manufacturer)	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety	Quality of life								
Kuna, 2011, Poland			X					X	X	X	X		X	X	X	X	X				3 yr	Alternaria alternata extract in a depot formulation with aluminum hydroxide/Novo-Helisen Depot® A alternata 100% (Allergopharma)	X	X	X	X	X	X	X	X	X	X	X	X	X							
Leynadier, 2001, France	X							X	X	X	X		X	X	X	X	X				1 yr	Standardized five-grass pollen (equal parts of: orchard, meadow, rye, sweet vernal and timothy) depot extract adsorbed onto calcium phosphate/Phostal® (Stallergenes)	X	X	X	X	X	X	X	X	X	X	X	X	X	X						
Metzger, 1981, England			X					X	X	X	X		X	X	X	X	X				5 wk	Glutaraldehyde-modified, tyrosine-adsorbed short ragweed extract/NR (Beecham Laboratories)	X													X						
Mirone, 2004, Italy			X					X	X	X	X		X	X	X	X	X				1 yr (DBRCT)	Ambrosia artemisiifolia adsorbed onto aluminum hydroxide and suspended in phenolated (0.4% w/v) saline solution/NR (ALK-Abelló)	X	X													X					
Olsen, 1995, Denmark	X	X	X					X	X	X	X		X	X	X	X	X				2 yr	Aluminum hydroxide-adsorbed extracts of standardized extracts of Betula, Phleum and Artemisia/Alutard® SQ (ALK)	X														X					
Ortolani, 1994, Italy			X					X	X	X	X		X	X	X	X	X				1 yr	Partially purified aiginate-conjugated extract of Parietaria judaica/Conjuvac Parietaria® (Dome Hollister-Stier)																X				
Pastorello, 1992, Italy	X							X	X	X	X		X	X	X	X	X				5-12 mo	Formalinized depot 6 grass allergoid adsorbed onto aluminum hydroxide/NR (Allergopharma)															X					
Patel, 2012, Canada						X		X	X	X	X		X	X	X	X	X				3 mo	Fel d 1-derived peptide antigen (Cat-PAD)/NR (Bachem and Patheon)	X <sup>a</sup>															X <sup>a</sup>				
Pauli, 2008, Austria, Denmark, France, Italy & Sweden		X						X	X	X	X		X	X	X	X	X				2 yr	Aluminum hydroxide-adsorbed vaccines of birch pollen extract, rBet v 1, and nBet v 1/NR (Stallergenes SA)	X	X															X			
Pfaar, 2010, Lithuania, Poland & Germany		X						X	X	X	X		X	X	X	X	X				19 mo	Standardized depigmented and glutaraldehyde-polymerized tree pollen extract (33% Corylus avellana, 33% Alnus glutinosa, 34% Betula alba) adsorbed onto aluminum hydroxide/Depigoid (Laboratorios LETI SL, Tres Cantos, Spain).	X																		X	
Pfaar, 2012, Germany	X							X	X	X	X		X	X	X	X	X				2 yr	Depigmented and glutaraldehyde-polymerized grass pollen mix adsorbed onto aluminum hydroxide/Depiquick® (Laboratorios LETI)	X	X																X	X	

(Continues)

TABLE 1 (Continued)

(a) Study (first author, y, country)	Allergen no.										Comparator					AIT Protocol					Product type/Name (manufacturer)				Short-term effectiveness		Long-term effectiveness		Safety	Quality of life					
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score							
Powell, 2007, UK Primary study Frew, 2006	X								X		X	X	X	X	X	X	X					14 mo										X			
Radcliffe, 2003, UK	X	X	X	X	X	X	X	X	X	X	X	X	X	X									2-3 mo		X							X			
Rak, 2001, Sweden	X								X		X	X	X	X								1 yr		X							X				
Riechelmann, 2010, Germany & Austria				X					X	X	X	X	X	X	X	X	X					1 yr		X	X	X	X	X	X	X	X	X	X		
Tabar, 2005, Spain				X					X		X	X	X	X	X	X	X					1 yr		X	X	X	X	X	X	X	X	X	X		
Tabar, 2008, Spain				X					X	X	X	X	X	X	X	X	X					18 mo		X	X	X	X	X	X	X	X	X	X	X	
Tari, 1997, Italy				X					X	X	X	X	X	X	X	X	X					2 yr		X	X	X	X	X	X	X	X	X	X	X	
Tworek, 2013, Poland	X	X	X	X					X	X	X	X	X	X	X	X	X					3 yr		X	X	X	X	X	X	X	X	X	X	X	
Varney, 1991, UK	X								X	X	X	X	X	X	X	X	X					8 mo		X	X	X	X	X	X	X	X	X	X	X	
Varney, 2003, UK					X				X	X	X	X	X	X	X	X	X					1 yr		X	X	X	X	X	X	X	X	X	X	X	
Walker, 2001, UK	X								X	X	X	X	X	X	X	X	X					2 yr		X	X	X	X	X	X	X	X	X	X	X	
Weyer, 1981, France	X								X	X	X	X	X	X	X	X	X					8 mo		X	X	X	X	X	X	X	X	X	X	X	
Zenner, 1997, Germany	X			X					X	X	X	X	X	X	X	X	X					4 mo		X	X	X	X	X	X	X	X	X	X	X	

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator			AIT Protocol					Short-term effectiveness		Long-term effectiveness		Quality of life								
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Other(s)	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrarush	Rx duration		Product type/Name (manufacturer)	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety
Ahmadiafshar, 2012, Iran	X							X	X	X	X		X	X							X	6 mo	10, 100, and 300 IR rye grass spray (Staloral 638)	X	X				X		
Alvarez-Cuesta, 2007, Spain					X			X	X	X	X											12 mo	Aqueous solution of cat dander extract with NaCl 0.9%, phenol 0.4% and glycerol 50% (protocol supplied by Laboratorios LETI, S.L.	X	X				X		
Amar, 2009, USA	X							X	X	X	X	X		X								10 mo	Monotherapy group: timothy extract Multiple allergen group: same amount of timothy plus 1 mL each of an additional 9 unstandardized extracts 1:20 wt/vol in 50% glycerin: maple, ash, juniper, American elm, cottonwood, Kochia, ragweed, sagebrush, and Russian thistle (ALK-Abelló).	X	X				X		
André, 2003, France			X					X	X	X	X			X								6.5 mo	Standardized ragweed extract (Stallergenes SA, Antony, France)	X	X				X		
Ariano, 2001, Italy & France		X						X	X	X	X			X								12 mo	Aqueous solution of an allergic fraction of <i>Cupressus arizonica</i> partially purified through dialysis in a physiological solution with 15% glycerin	X	X				X		
Aydogan, 2013, Turkey, UK & Cyprus.					X			X	X	X	X											12 mo	1:1 mixture of <i>D. pteronyssinus</i> and <i>D. farinae</i> (STALORAL, Stallergenes SA, Antony, France)	X	X				X		
Bahçeciler, 2007, Turkey					X			X	X	X	X											6 mo	<i>D. pteronyssinus</i> and <i>D. farinae</i> 50/50 extract.	X	X				X		
Bergmann, 2013, Germany, France, the Netherlands & Spain					X			X	X	X	X											2 yr	Oral tablets of 1:1 mixture of <i>D. pteronyssinus</i> and <i>D. farinae</i> (28 mg and 120 mg respectively for the 500 IR tablet, 16 mg and 68 mg respectively for the 300 IR tablet)	X	X			X	X		
Blaiss, 2010, USA & Canada	X							X	X	X	X											18 mo	f 2800 bioequivalent allergen units of grass AIT treatment (oral lyophilisate, <i>Phileum pratense</i> , 75 000 standardized quality tablet, containing approximately 15 mg of Phl p 5; Schering-Plough Corp, a division of Merck & Co, Kenilworth, NJ)	X	X			X	X		X
Bowen, 2004, Canada			X					X	X	X	X			X								4 mo	Ragweed allergen extract	X	X				X		
Bozek, 2012, Poland		X						X	X	X	X											3 yr	Oral Staloral 300 SR Der p and Der f (1:1)	X	X				X		

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator		AIT Protocol					Product type/Name (manufacturer)		Short-term effectiveness		Long-term effectiveness		Quality of life							
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Symptom score		Medication score	Combined score	Symptom score	Medication score	Combined score	Safety	
Bozek, 2014, Poland	X							X		X												3 yr	X	X					X		
Bufe, 2004, Germany	X							X		X			X									3 yr	X	X					X		
Bufe, 2009, Germany	X							X		X			X									8-23 wk	X	X					X		
Caffarelli, 2000, Italy	X							X		X			X									3 mo	X	X					X		
Clavel, 1998, France	X							X		X												7 mo	X	X					X		
Cortellini, 2010, Italy			X					X		X												10 mo	X	X					X		
Cox, 2012, USA	X							X		X												6 mo	X	X	X	X	X	X	X	X	X
Creticos, 2013, USA			X					X		X			X									20 mo	X	X	X	X	X	X	X	X	X
Creticos, 2013, Canada			X					X		X			X									12 wk	X	X					X		X
Dahl, 2006, Denmark, Germany, Italy, the Netherlands, Sweden, Austria, Spain & UK	X							X		X			X									1 yr	X	X	X	X	X	X	X	X	X

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator		AIT Protocol							Short-term effectiveness		Long-term effectiveness		Quality of life						
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Product type/Name (manufacturer)		Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score
Dahl, 2006, Denmark & Sweden	X							X		X				X	X								Orodispersible grass allergen tablet (Grazax; approximately 15 mg major allergen <i>Phleum pratense</i> (75 000 SQ-T)	X	X					X
de Blay, 2007, France	X							X		X				X	X							12 mo	3-grass pollen extract (33.3% <i>Dactylis glomerata</i> [orchard grass], 33.3% <i>Phleum pratense</i> timothy grass), and 33.3% <i>Lolium perenne</i> [rye grass]) Allerbio, Varennes-en-Argonne, France) in 50% glycerin	X	X					X
De Bot, 2011, the Netherlands					X			X		X												2 yr	Aqueous extract of house dust mites ( <i>D. pter</i> ) in a glycerinated isotonic phosphate-buffered solution (Oralgen Mijten)/placebo treatment consisting of the glycerol-containing solvent	X	X					X
Demoly, 2015, Europe					X			X		X												1 yr	1:1 mixture of two species of house dust mite allergens ( <i>D. pteronyssinus</i> and <i>D. farinæ</i> ) (1:1:1:1 ratio of the major allergens <i>Der p 1</i> , <i>Der f 1</i> , <i>Der p 2</i> , and <i>Der f 2</i> )	X	X					X
Didier, 2007, Europe								X		X				X	X							6 mo	Mixture of 5 grass pollens (orchard, meadow, perennial rye, sweet vernal, and timothy grasses)	X						X
Didier, 2009, France, Germany & Spain	X							X		X				X	X							6 mo	Lyophilized vaccines of five grass pollens (orchard or cocksfoot ( <i>Dactylis glomerata</i> ), meadow ( <i>Poa pratensis</i> ), perennial rye ( <i>Lolium perenne</i> ), sweet vernal ( <i>Anthoxanthum odoratum</i> ), and timothy ( <i>Phleum pratense</i> ))	X						X
Didier, 2013, Denmark, Austria, France, Canada & Germany	X							X		X				X	X							4 yr	300IR tablets containing mixture of 5 grasses [cocksfoot ( <i>Dactylis glomerata</i> ), meadow ( <i>Poa pratensis</i> ), rye ( <i>Lolium perenne</i> ), sweet vernal ( <i>Anthoxanthum odoratum</i> ) and timothy ( <i>Phleum pratense</i> )	X						X
Durham, 2005, Canada, Denmark & Sweden	X							X		X				X	X							2 yr	Fast-dissolving grass allergen tablet (ALK-Abelló A/S) containing timothy grass extract ( <i>Phleum pratense</i> )	X	X					X
Durham, 2007, UK Primary study: Dahl, 2006	X							X		X				X	X							16 wk	Grass allergen tablet (Grazax)	X						X

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator			AIT Protocol				Short-term effectiveness		Long-term effectiveness		Quality of life																				
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash		Rx duration	Product type/Name (manufacturer)	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety											
Durham, 2009, UK	X							X		X	X		X	X								3 yr	Grass allergen tablet with <i>Phleum pratense</i> 75 000 SQ-T/2800 BAU (ALK-Abelló, Hørsholm, Denmark) (Grazax)																			
Results after 1-y follow-up of the Dahl (2006) study																																										
Durham, 2011, UK	X							X		X	X		X	X								2 yr	SQ-standardized grass allergy tablet ( <i>Phleum pratense</i> 75 000 SQ-T/2800 BAU, ALK, Denmark) (Grazax)																			
Results of 2-y follow-up of the Dahl (2006) trial																																										
Durham, 2012, UK, Austria, Germany, the Netherlands, Sweden & Denmark	X							X		X	X		X	X								3 yr	SQ-standardized grass allergy tablet (Grazax)																			
Results of 2-y follow-up of the Dahl (2006) trial																																										
Drachenberg, 2002, Germany	X							X		X	X		X	X										Grass, rye or birch pollens																		
Feliziani, 1995, Italy	X							X		X	X		X	X										Grass pollen extracts (5 x 1 drop of 0.04 BU/mL, up until 5 x 1 drop of 100 BU/mL)																		
Frølund, 2010, Austria, Denmark & UK	X							X		X	X		X	X								4 yr	SQ-standardized grass allergy immunotherapy tablet (AIT), Grazax ( <i>Phleum pratense</i> 75 000 SQ-T/2800 BAU, ALK, Denmark).																			
Guez, 2000, France	X							X		X	X		X	X								24 mo	<i>D. pteronyssinus</i> and <i>D. farinæ</i> 50/50 extract																			
Halken, 2010, Germany, Denmark, Poland, France & Spain	X							X		X	X		X	X								10 mo	Five-grass pollen 300IR tablets (Stallergenes SA, France)																			
Hirsch, 1997, Germany	X							X		X	X		X	X								12 mo	Purified <i>D. pteronyssinus</i> extract in 50% aqueous glycerol (cumulative dose 570 jag) (Allergopharma J. Ganzer KG, Reinheke, FRG)																			
Horak, 1998, Austria	X							X		X	X		X	X								4 mo	Biologically standardized <i>Betula Alba</i> Alergia e Immunologia Abello SA																			
Horak, 2009, Austria	X							X		X	X		X	X								4 mo	300-IR 5-grass pollen tablet (orchard, meadow, perennial rye, sweet vernal, timothy)																			

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator				AIT Protocol				Product type/Name (manufacturer)		Short-term effectiveness		Long-term effectiveness		Quality of life				
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrash	Rx duration	Symptom score	Medication score		Symptom score	Medication score	Combined score	Safety
Hordijk, 1998, the Netherlands	X							X		X			X									10 mo	X					X	
Ibanez, 2007, Spain & Germany	X							X														28 d						X	
Ippoliti, 2003, Italy					X			X		X												6 mo						X	
Kaluzinska-Parzyzek, 2011, Poland (Polish, translated)	X							X		X								X				2 yr							
La Rosa, 1999, Italy & France			X					X		X												2 yr					X	X	
Marcucci, 2003, Italy					X			X		X												1 yr							
Moreno-Ancillo, 2007, Spain	X								X	X												10 mo					X	X	X
Mosbeck, 2014, Denmark, Italy, Germany & France					X			X		X												1 yr					X	X	
Mosges, 2007, Germany	X								X	X												9 mo					X	X	X
Okubo, 2008, Japan		X						X		X												7 mo					X	X	X

(Continues)



**TABLE 1** (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator		AIT Protocol			Short-term effectiveness			Long-term effectiveness																											
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others)	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Coseseasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrarush	Rx duration	Product type/Name (manufacturer)	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Symptom score	Medication score	Combined score	Safety	Quality of life												
Ott, 2009, Germany	X							X	X	X	X	X	X								X	3 yr	Pollen extract mixture of five grasses (cocksfoot or orchard, meadow, perennial rye, sweet vernal and timothy grasses; Staloral, Stallergenes SA, France) (300 IR/mL, equivalent to 21 lg/mL of Phleum pratense major allergen)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X						
Nelson, 1993, USA					X			X	X	X	X	X	X									105 d	Cat dander extract (total dose: 4.5 AU)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X							
Pajno, 2003, Italy			X					X	X	X	X	X	X									14 mo	<i>P. judaica</i> , fluticasone	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X							
Palma-Carlos, 2006, Italy	X							X	X	X	X	X	X									2 yr	Mixture of carbamylated grass pollens ( <i>Holcus lanatus</i> 33%, <i>Phleum pratense</i> 33%, and <i>Poa pratensis</i> 33%) in tablets	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X			
Panzner, 2008, Czech Republic	X							X	X	X	X	X	X									1 yr	Mixture of six grass pollen species extracts (oat grass ( <i>Arrhenatherum elatius</i> ), orchard grass ( <i>Dactylis glomerata</i> ), fescue ( <i>Festuca sp.</i> ), rye grass ( <i>Lolium sp.</i> ), timothy grass ( <i>Phleum pratense</i> ), and rye ( <i>Secale cereale</i> )) (H-AI per os) (Sevapharma A.S., Prague, Czech Republic)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Passalacqua, 1996, Italy					X			X	X	X	X	X	X									2 yr	Monomeric allergoid tablets with <i>Dermatophagoides pteronyssinus</i> and <i>D farina</i>	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X			
Passalacqua, 1999, Italy	X							X	X	X	X	X	X									7 mo	ALK-Abelló (major allergen Par j) (0.016, 0.08, 0.4, 2, and 10 BU/mL)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X				
Passalacqua, 2006, Italy					X			X	X	X	X	X	X									2 yr	Monomeric carbamylated grass pollen allergen (Lais)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X					
Pfaar, 2008, Germany, Poland & Macedonia	X							X	X	X	X	X	X									2 yr	Six-grass pollen mixture (high-dose)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X				
Pradallier, 1999, France	X							X	X	X	X	X	X									4.5 mo	Five-grass pollen extracts (orchard grass, meadow grass, ryegrass, sweet vernal grass, and timothy grass) (Stallergenes SA, Antony, France)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X			
Purello-D'Ambrosio, 1999, Italy		X						X	X	X	X	X	X									6 mo	<i>P. judaica</i> extract (five 3-ml vials: 0.016 BU/ml (vial 0), 0.08 (#1), 0.04 (#2), 2.00 (#3), and 10.00 (#4) in physiological saline with 50% v/v of glycerol & 0.4% w/v of phenol) (maximum concentration of major allergen Par j 1: 0.6 mg/mL)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	

(Continues)

TABLE 1 (Continued)

(b) Study (first author, y, country)	Allergen no.										Comparator				AIT Protocol				Product type/Name (manufacturer)				Short-term effectiveness		Long-term effectiveness		Quality of life		
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Other(s)	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster	Semirush	Rush	Ultrarush	Rx duration	Symptom score	Medication score	Combined score	Symptom score		Medication score	Combined score
Queiros, 2013, Brazil & USA					X			X	X	X	X	X	X									18 mo			X				X
Rak, 2006, UK	X							X	X	X	X											174 d							X
Roder, 2007, the Netherlands	X							X	X	X	X				X							2 yr		X	X				X
Rolinck-Werninghaus, 2004, Germany	X							X	X	X	X				XX							32 mo		X	X				X
Sabbah, 1994, France	X							X	X	X	X											4 mo		X	X				X
Stelmach, 2011, Poland	X							X	X	X	X											2 yr		X	X				X
Tari, 1990, Italy					X			X	X	X	X											18 mo		X					X
Valovirta, 2006, Finland	X							X	X	X	X											19 mo		X	X				X
Van Niekerk, 1987, South Africa	X							X	X	X	X											24 mo		X					X
Vourdas, 1998, Greece & France	X							X	X	X	X											2 yr		X	X				X

(Continues)

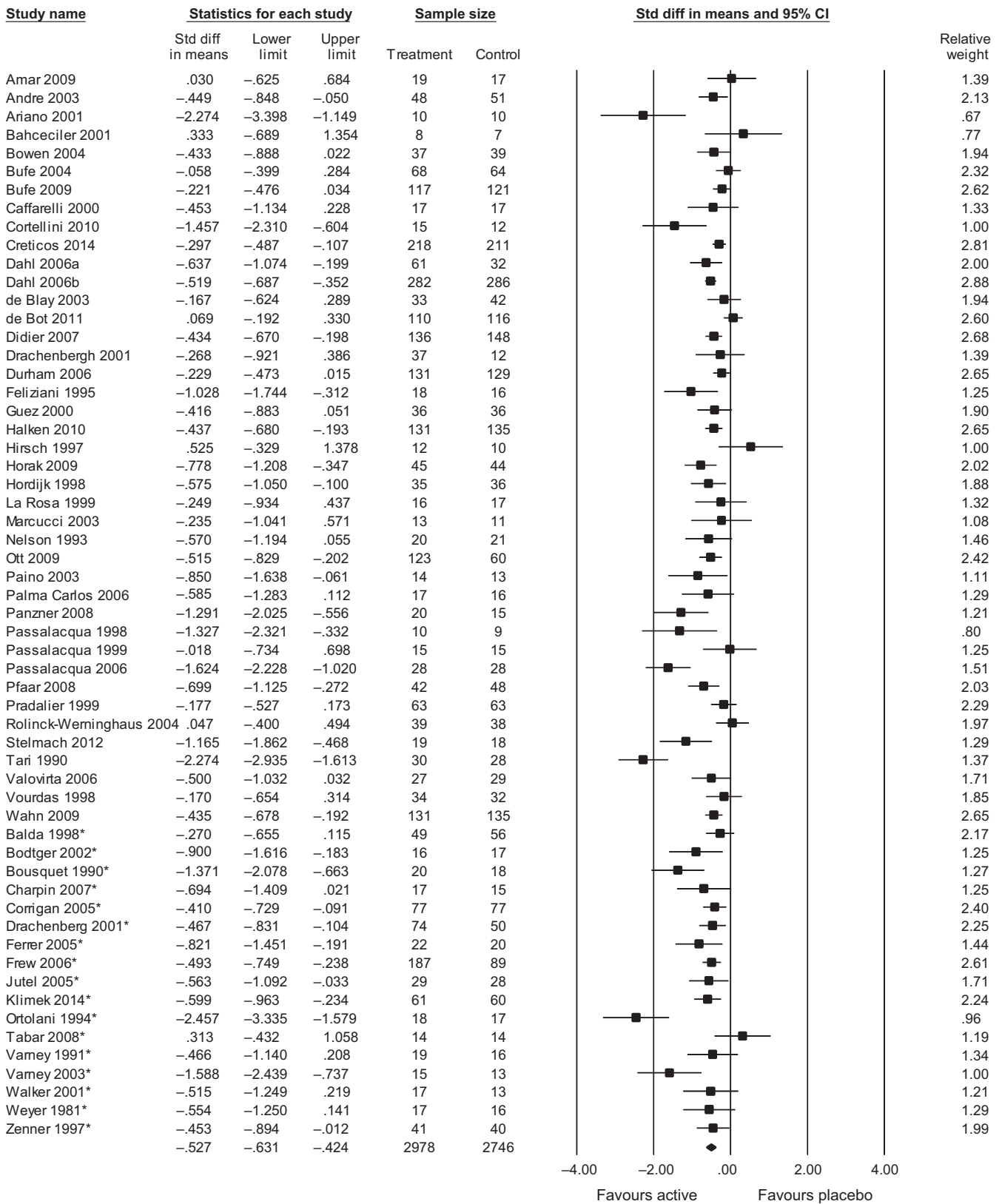
TABLE 1 (Continued)

(b) Study (first author, Y, country)	Allergen(s) type		Allergen no.		Comparator				AIT Protocol				Product type/Name (manufacturer)		Short-term effectiveness		Long-term effectiveness		Quality of life												
	Grass pollen(s)	Tree pollen(s)	Weed(s)	Mold(s)	House dust mite	Cat	Dog	Others	Single	Multiple	Placebo	Routine care	Active	Preseasonal	Cosasonal	Continuous	Conventional	Cluster		Semirush	Rush	Ultrush	Rx duration	Symptom score	Medication score	Symptom score	Medication score	Combined score	Safety		
Wang, 2013, China	X				X			X			X											6 mo	X	X	X	X	X	X			
Wahn, 2012, Germany & Poland	X				X			X			X											8 mo	X	X	X	X	X	X			
Wahn, 2009, Denmark & France	X				X			X			X											Approx 5-6 mo	X	X	X	X	X	X			
(c) Study (first author, year, country)	Allergen(s) type		Allergen number		Route AIT		Comparator		AIT Protocol				Product type/Name (manufacturer)		Short-term effectiveness		Long-term effectiveness		Quality of life												
	Grass pollen(s)	Tree pollen(s)	Weeds	Molds	House dust mite	Cat	Dog	Others	Single	Multiple	SCIT	SLIT	ILIT	Placebo	Routine care	Active	Preseasonal	Cosasonal		Continuous	Conventional	Cluster	Semirush	Rush type	Ultrush	Duration of Rx	Symptom score	Medication score	Symptom score	Medication score	Combined score
Hylander et al., 2016, Spain	X	X			X				X		X	X	X	X			X							2 mo	X	X	X	X	X	X	
Senti et al., 2012, Switzerland					X				X		X	X	X	X										2 mo	X <sup>b</sup>	X <sup>b</sup>	X <sup>b</sup>	X <sup>b</sup>	X <sup>b</sup>	X <sup>b</sup>	X <sup>b</sup>

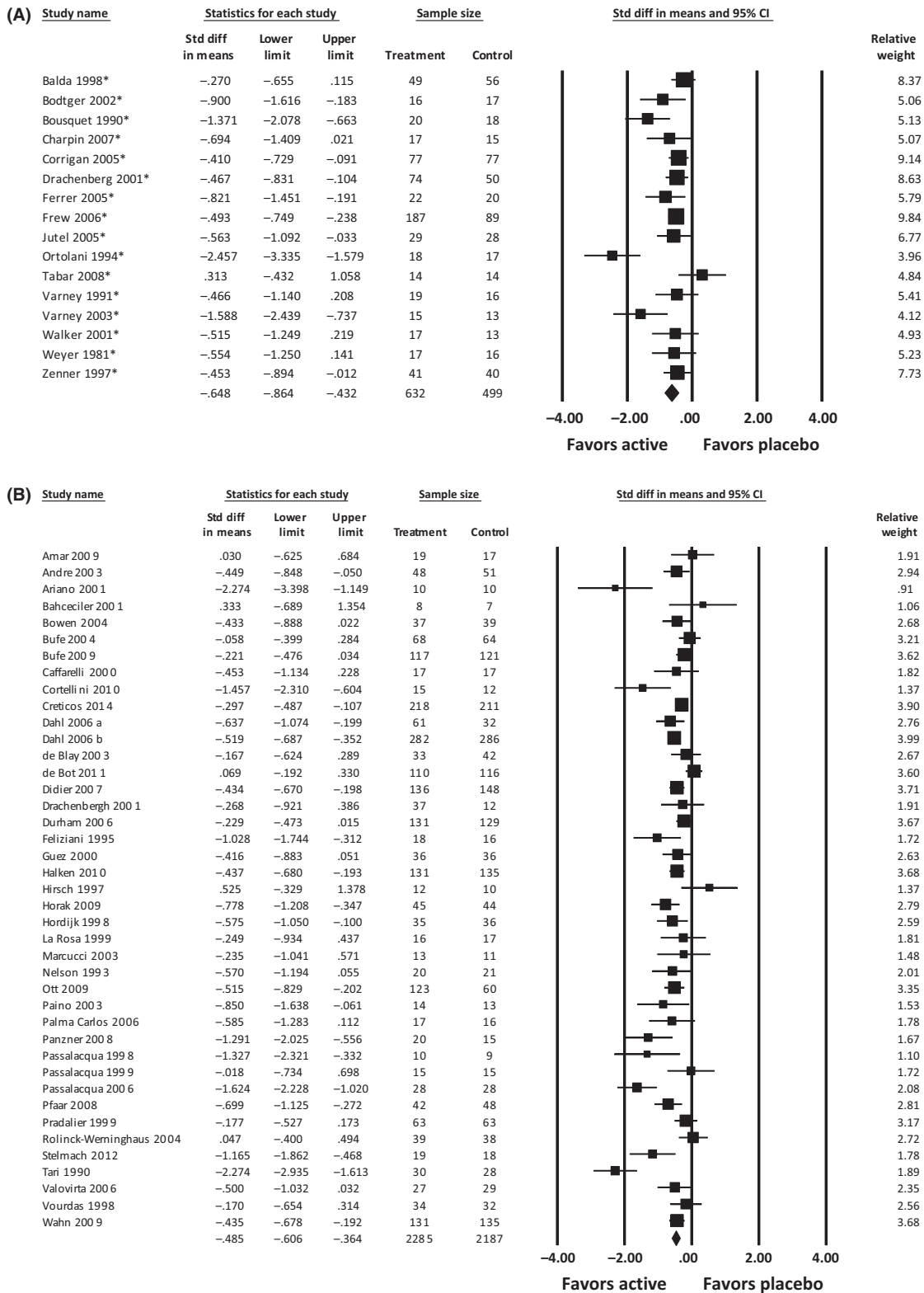
AIT, allergen-specific immunotherapy; ILIT, intralymphatic immunotherapy; mo, month; NBS, not better specified; NR, not reported; Rx, treatment; SCIT, subcutaneous immunotherapy; SLIT, sublingual immunotherapy; UR, unclear reporting wk, week; yr, year.

<sup>a</sup>Environmental exposure chamber.

<sup>b</sup>Assessment after 300 days of discontinuation of ILIT.

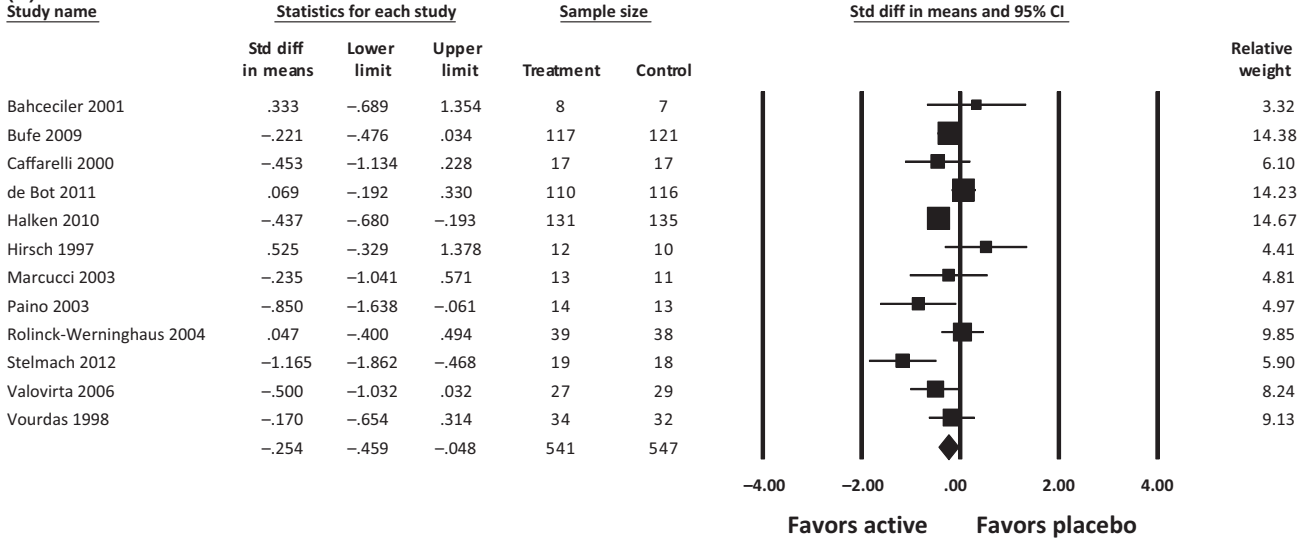


**FIGURE 2** Meta-analysis of double-blind RCTs comparing symptom scores between allergen immunotherapy (AIT) (subcutaneous immunotherapy (SCIT) or sublingual immunotherapy (SLIT)) and placebo groups (random-effects model). Heterogeneity:  $\tau^2=.090$ ;  $\chi^2=173.586$ ,  $df=57$  ( $P<.0001$ );  $I^2=67\%$ . Test for overall effect:  $Z=-9.992$  ( $P<.0001$ ). \*denotes SCIT studies

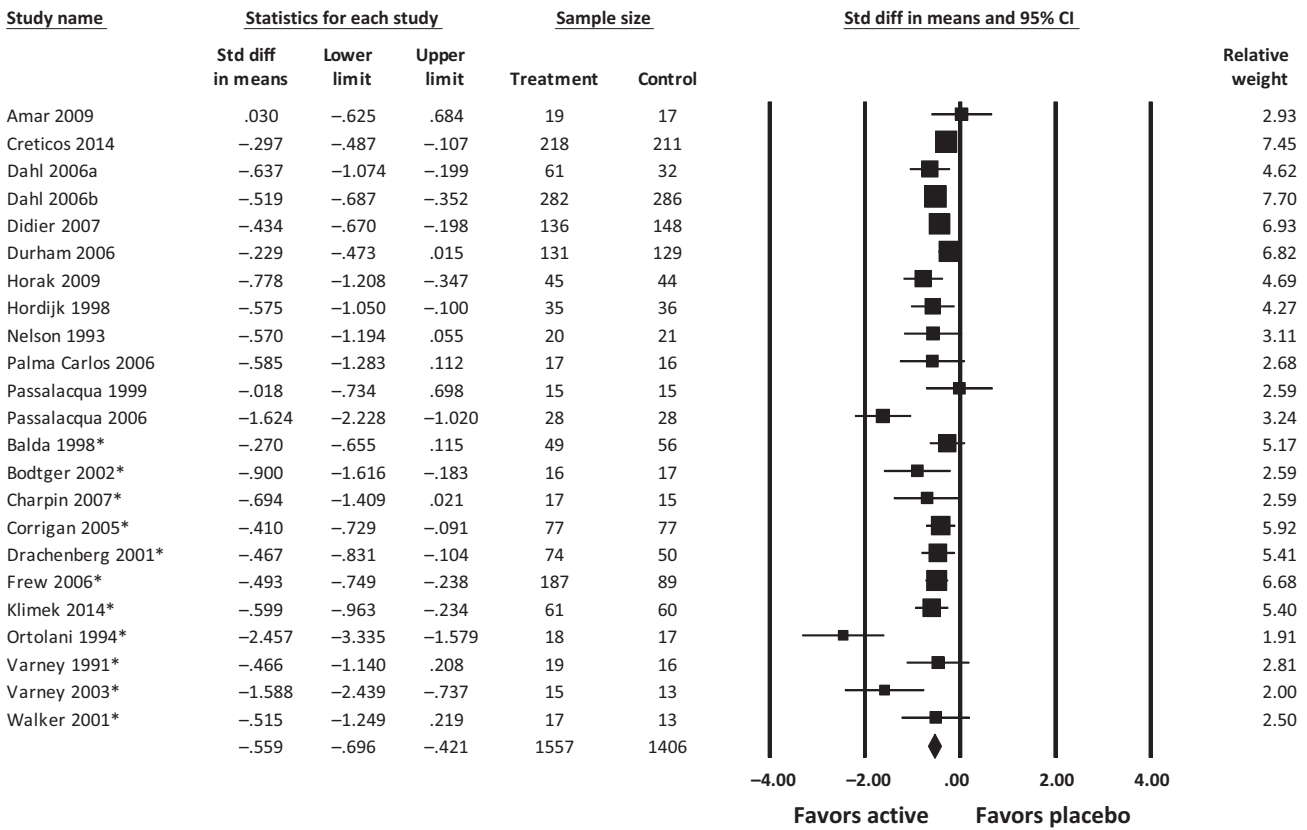


**FIGURE 3** Meta-analysis of double-blind RCTs comparing symptom scores between (A) subcutaneous immunotherapy (SCIT) and placebo groups and (B) sublingual immunotherapy (SLIT) and placebo group (random-effects models). (A) Heterogeneity:  $\tau^2=.106$ ;  $\chi^2=39.357$ ,  $df=15$  ( $P<.001$ );  $I^2=62\%$ . Test for overall effect:  $Z=-5.875$  ( $P<.0001$ ). \*denotes SCIT studies. (B) Heterogeneity:  $\tau^2=.088$ ;  $\chi^2=129.171$ ,  $df=40$  ( $P<.0001$ );  $I^2=69\%$ . Test for overall effect:  $Z=-7.855$  ( $P<.0001$ ).

(A)



(B)



**FIGURE 4** Meta-analysis of double-blind RCTs comparing symptom scores between allergen immunotherapy (AIT) (subcutaneous immunotherapy (SCIT) or sublingual immunotherapy (SLIT)) and placebo group in (A) those <18 years old and (B) those ≥ 18 years old (random-effects models). (A) Heterogeneity:  $\tau^2=.059$ ;  $\chi^2=24.209$ ,  $df=11$  ( $P<.012$ );  $I^2=54\%$ . Test for overall effect:  $Z=-2.423$  ( $P<.015$ ). (B) Heterogeneity:  $\tau^2=.057$ ;  $\chi^2=57.748$ ,  $df=22$  ( $P<.0001$ );  $I^2=62\%$ . Test for overall effect:  $Z=-7.969$  ( $P<.0001$ ). \*denotes SCIT studies

- Seasonal vs perennial allergens for SCIT: SMD  $-0.49$  (95% CI  $-0.72, -0.27$ ) for seasonal and SMD  $-1.59$  (95% CI  $-2.44, -0.74$ ) for perennial (results from only one study) (Figures S5A and B), these demonstrating evidence of benefit from both approaches.
- Seasonal vs perennial allergens for SLIT: SMD  $-0.35$  (95% CI  $-0.45, -0.26$ ) for seasonal and SMD  $-0.81$  (95% CI  $-1.41, -0.20$ ) for perennial allergens (Figure S6A,B)
- Pre-/coseasonal vs continuous treatment in SCIT for pollen: SMD  $-0.51$  (95% CI  $-0.63, -0.38$ ) in pre-/coseasonal and SMD  $-0.69$

(95% CI  $-1.09, -0.29$ ) (Figures S7A and B), these analyses demonstrating evidence of benefit from both approaches.

- Pre-/coseasonal vs continuous treatment in SLIT for pollens: SMD  $-0.40$  (95% CI  $-0.48, -0.32$ ) in pre-/coseasonal and SMD  $-0.55$  (95% CI  $-0.98, -0.11$ ) in continuous (Figures S8A and B), these analyses demonstrating a clear benefit associated with both approaches.
- Modified allergen extracts (allergoids) vs unmodified allergen extracts in SCIT: SMD  $-0.60$  (95% CI  $-0.89, -0.31$ ) vs SMD  $-0.65$  (95% CI  $-0.93, -0.36$ ) (Figures S9A and B), these analyses demonstrating evidence of benefit from both modalities
- Aqueous solutions vs tablets in SLIT: SMD  $-0.42$  (95% CI  $-0.68, -0.15$ ) in aqueous and SMD  $-0.53$  (95% CI  $-0.73, -0.34$ ) with tablets (Figures S10A and B), these analyses confirming benefit with both preparations.
- Different allergens for AIT (SCIT and SLIT): HDM: SMD  $-0.73$  (95% CI  $-1.37, -0.10$ ); grass: SMD  $-0.45$  (95% CI  $-0.54, -0.36$ ); tree: SMD  $-0.57$  (95% CI  $-0.92, -0.21$ ); molds: SMD  $-0.56$  (95% CI  $-2.29, 1.18$ ); weeds: SMD  $-0.68$  (95% CI  $-1.06, -0.30$ ), these showing that AIT was clearly effective for all allergens except molds for which there was evidence suggestive of benefit but this was imprecisely estimated (Figure S11A-E),

### Long-term

To investigate long-term effectiveness, a number of investigators studied a discontinuation period following trials that involved randomization to AIT or placebo in which the superiority of AIT was confirmed. In this longer-term phase, patients were followed up and outcomes were then again assessed at least one year post-discontinuation of AIT.

There were four trials that studied this outcome, one SCIT<sup>42</sup> and three SLIT,<sup>89,114,133</sup> all of which were judged to be at low ROB. Meta-analysis of data was not possible. A full descriptive summary of the main findings are provided in the supplement. In summary, all four trials at low ROB found a beneficial effect on the long-term effectiveness of AIT on symptom scores.

## 3.2.2 | Medication scores

### Short-term

Eighty nine studies reported on the short-term effectiveness of AIT administered by the SCIT ( $n=46$ ), SLIT ( $n=42$ ) and ILIT ( $n=1$ ) routes on medication scores.

We were able to pool data from 45 SCIT and SLIT trials. This showed an overall SMD of  $-0.38$  (95% CI  $-0.49, -0.26$ ), this suggesting a small-to-medium effect in favor of AIT in improving medication scores (Figure 5).

**Sensitivity analyses** Sensitivity analysis, performed by excluding all studies at high ROB, gave an SMD of  $-0.35$  (95% CI  $-0.46, -0.24$ ) (Figure S12).

**Assessment of publication bias** The Funnel plot revealed evidence of potential publication bias (Figure S13) which was also suggested by the Begg ( $P=0.004$ ) and Egger ( $P=0.03$ ) tests.

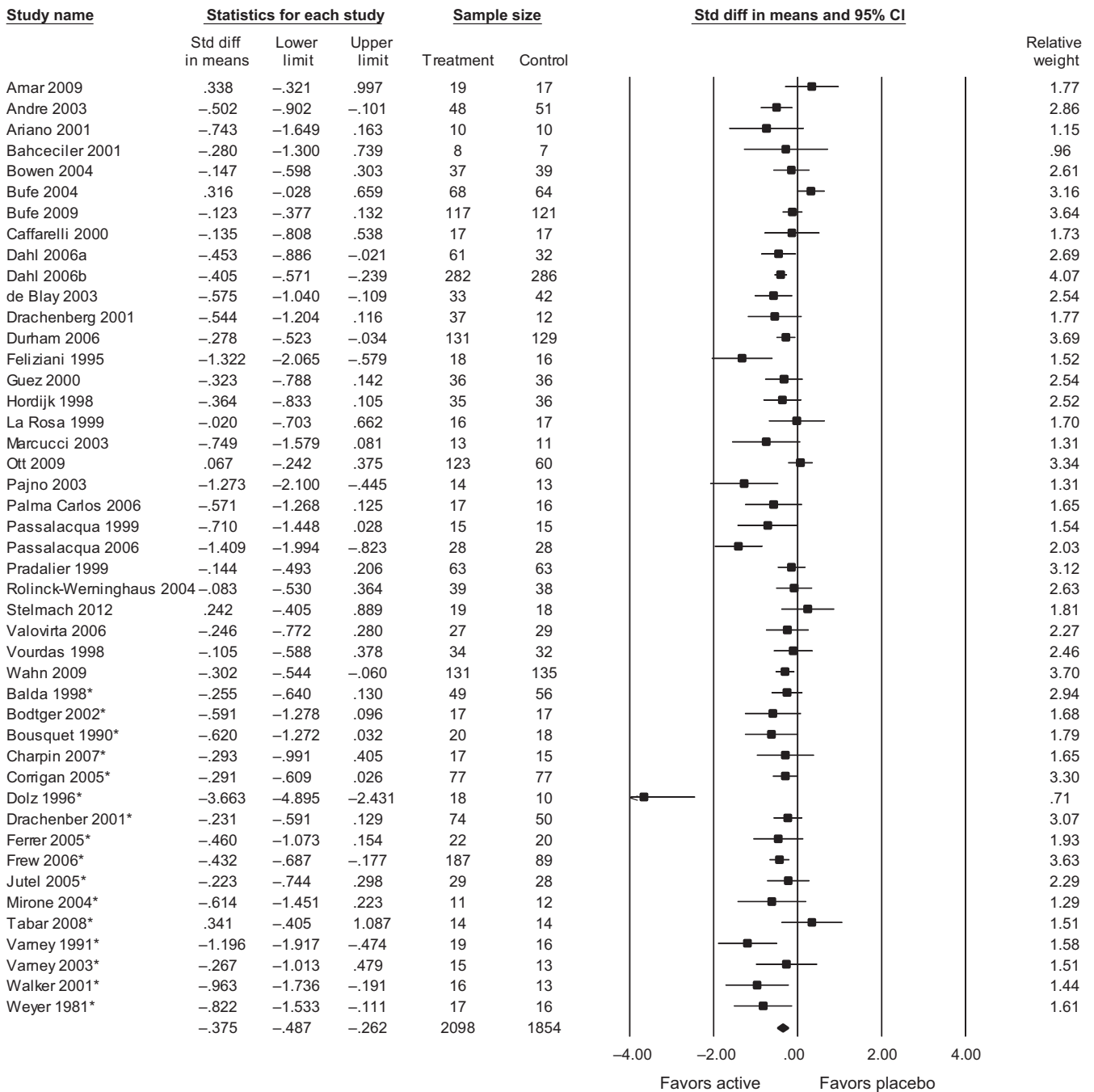
**Subgroup analyses** Subgroup analyses were undertaken to compare:

- SCIT vs SLIT: SMD  $-0.52$  (95% CI  $-0.75, -0.29$ ) for SCIT and  $-0.31$  (95% CI  $-0.44, -0.18$ ) for SLIT (Figures 6A and B), these analyses demonstrating that both routes were effective.
- Children vs adults: SMD  $-0.21$  (95% CI  $-0.42, 0.01$ ) for children and SMD  $-0.43$  (95% CI  $-0.56, -0.30$ ) for adults (Figures S14A and B), these showing a clear benefit in adults and the suggestion of benefit in children (but this was not confirmed)
- Children vs adults for SLIT only: SMD  $-0.60$  (95% CI  $-1.12, -0.07$ ) for children and SMD  $-0.45$  (95% CI  $-0.69, -0.22$ ) for adults showing a benefit in both (Figures S15A and B).
- Seasonal vs perennial allergens for AIT (SCIT and SLIT): SMD  $-0.30$  (95% CI  $-0.43, -0.16$ ) for seasonal and SMD  $-0.63$  (95% CI  $-1.12, -0.15$ ) for perennial allergens (Figures S16A and B), these indicating that both were effective.
- Seasonal vs perennial allergens for SCIT: SMD  $-0.77$  (95% CI  $-1.28, -0.25$ ) for seasonal and SMD  $-0.27$  (95% CI  $-1.01, 0.48$ ) for perennial (results from only one study) (Figures S17A and B)
- Seasonal vs perennial allergens for SLIT: SMD  $-0.24$  (95% CI  $-0.38, -0.10$ ) for seasonal, SMD  $-0.72$  (95% CI  $-1.30, -0.13$ ) (Figures S18A and B), indicating that both were effective.
- Pre-/coseasonal vs continuous treatment in SCIT for pollens: SMD  $-0.40$  (95% CI  $-0.56, -0.25$ ) in pre-/coseasonal and SMD  $-1.23$  (95% CI  $-2.34, -0.12$ ) in continuous (Figures S19A and B), these indicating that both were effective.
- Pre-/coseasonal vs continuous treatment in SLIT for pollens: SMD  $-0.30$  (95% CI  $-0.42, -0.18$ ) in pre-/coseasonal and SMD  $0.00$  (95% CI  $-0.32, 0.33$ ) for continuous (Figures S20A and B), these analyses suggesting that pre-/coseasonal was effective and that continuous treatment was ineffective.
- Modified allergen extracts (allergoids) vs unmodified allergen extracts in SCIT SMD  $-0.94$  (95% CI  $-1.73, -0.16$ ) vs SMD  $-0.44$  (95% CI  $-0.64, -0.24$ ) (Figures S21A and B),
- Aqueous solutions vs tablets in SLIT: SMD  $-0.42$  (95% CI  $-0.68, -0.15$ ) for those receiving aqueous and SMD  $-0.53$  (95% CI  $-0.73, -0.34$ ) for tablets (Figures S22A and B), these analyses showing that both preparations were effective.
- Different allergens for AIT (SCIT and SLIT): HDM: SMD  $-0.63$  (95% CI  $-1.12, -0.15$ ) vs Grass: SMD  $-0.32$  (95% CI  $-0.46, -0.18$ ) vs Tree: SMD  $-0.40$  (95% CI  $-0.59, -0.20$ ) vs Molds: SMD  $0.34$  (95% CI  $-0.41, 1.09$ )(results from only one study) vs Weeds: SMD  $-0.44$  (95% CI  $-0.80, -0.09$ ) (Figures S23A-E), these showing evidence of benefit for all allergens except molds.

### Long-term

There were three low ROB trials that assessed this outcome: one SCIT<sup>42</sup> and two SLIT.<sup>114,133</sup> These three trials are described in detail





**FIGURE 5** Meta-analysis of double-blind RCTs studies comparing medication scores between allergen immunotherapy (AIT) (subcutaneous immunotherapy (SCIT) or sublingual immunotherapy (SLIT)) and placebo groups (random-effects model). Heterogeneity:  $\tau^2=.074$ ;  $\chi^2=110.337$ ,  $df=44$  ( $P<.0001$ );  $I^2=60\%$ . Test for overall effect:  $Z=-6.502$  ( $P<.0001$ ). \*denotes SCIT studies

in the supplement. Overall, one trial found a benefit of AIT (SCIT) on long-term medication scores; the two other SLIT trials did not show a sustained effect.

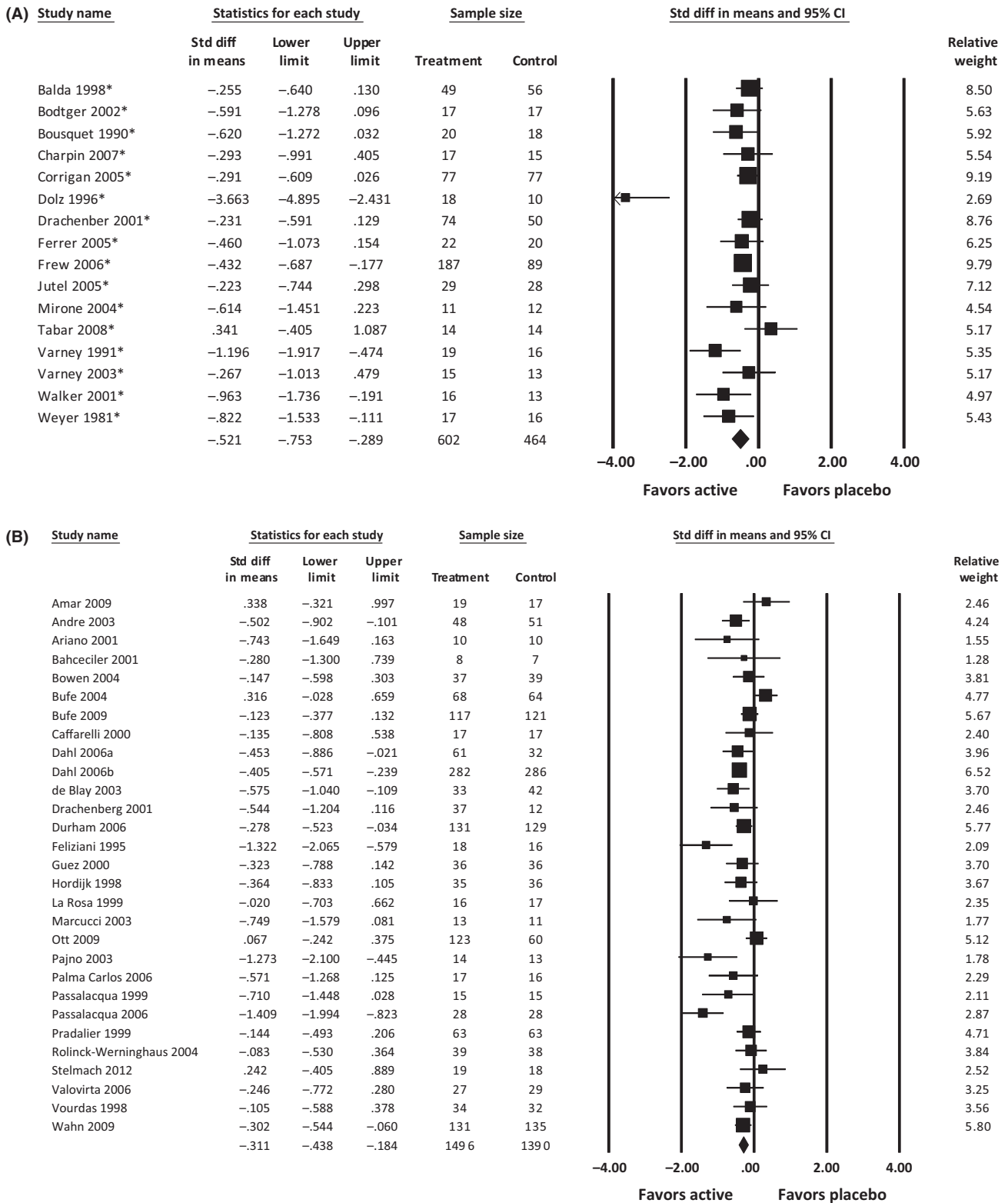
**3.2.3 | Combined symptom and medication scores**

Twenty-nine studies reported on the short-term effectiveness of AIT administered by the SCIT (n=20) and SLIT (n=9) routes on combined

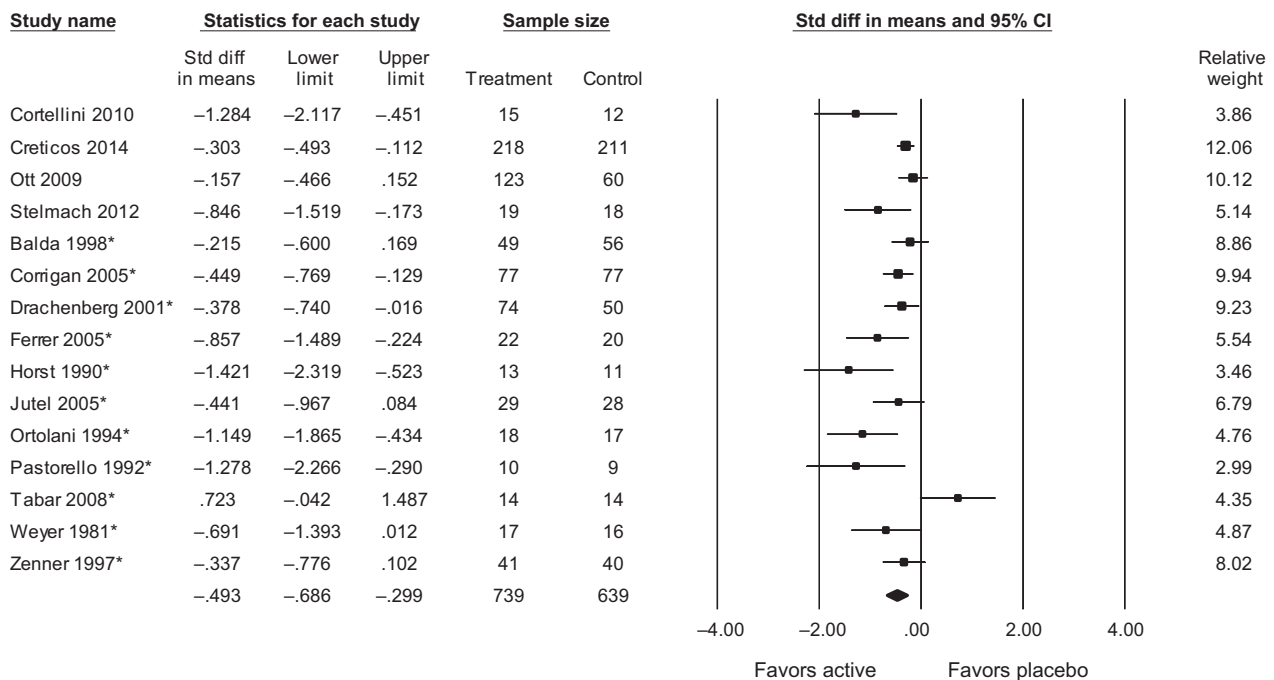
symptom and medication scores. Two studies (one SCIT and one SLIT) reported on long-term effectiveness in relation to this outcome.

**Short-term**

We were able to pool data from 15 studies. Meta-analysis found a SMD of  $-0.49$  (95% CI  $-0.69, -0.30$ ), this suggesting a small-to-moderate effect in favor of AIT (Figure 7).



**FIGURE 6** Meta-analysis of double-blind RCTs comparing medication scores between (A) subcutaneous immunotherapy (SCIT) and placebo groups and (B) sublingual immunotherapy (SLIT) and placebo groups (random-effects models). (A) Heterogeneity:  $\tau^2=.126$ ;  $\chi^2=42.241$ ,  $df=15$  ( $P<.0001$ );  $I^2=64\%$ . Test for overall effect:  $Z=-4.399$  ( $P<.0001$ ). \*denotes SCIT studies. (B) Heterogeneity:  $\tau^2=.057$ ;  $\chi^2=64.535$ ,  $df=28$  ( $P<.0001$ );  $I^2=57\%$ . Test for overall effect:  $Z=-4.805$  ( $P<.0001$ )



**FIGURE 7** Meta-analysis of double-blind RCTs studies comparing combined symptom and medication scores between allergen immunotherapy (AIT) (subcutaneous immunotherapy (SCIT) or sublingual immunotherapy (SLIT)) and placebo groups (random-effects model). Heterogeneity:  $\tau^2=.071$ ;  $\chi^2=33.631$ ,  $df=14$  ( $P<.002$ );  $I^2=58\%$ . Test for overall effect:  $Z=-4.997$  ( $P<.001$ ). \*denotes SCIT studies

**Sensitivity analysis** No sensitivity analysis was possible as no studies were judged to be at high ROB.

**Publication bias** The funnel plot showed evidence of potential publication bias, (Figure S24) which was also suggested by the Begg ( $P=0.005$ ) and Egger ( $P=0.03$ ) tests.

**Subgroup analyses** Subgroup analyses were undertaken to compare:

- SCIT vs SLIT: SMD  $-0.51$  (95% CI  $-0.77, -0.26$ ) for SCIT and SMD  $-0.47$  (95% CI  $-0.81, -0.12$ ) (Figures 8A and B), these analyses showing a benefit from both SCIT and SLIT.
- Children ( $<18$ ) vs adults ( $\geq 18$  years) for AIT (SCIT and SLIT): SMD  $-0.85$  (95% CI  $-1.52, -0.17$ ) (results from one study only) for children and SMD  $-0.44$  (95% CI  $-0.65, -0.22$ ) for adults (Figures S25A and B), these analyses showing a benefit in both children and adults
- Pre-/coseasonal (short-term treatment) vs continuous treatment in SCIT for pollen: SMD  $-0.41$  (95% CI  $-0.58, -0.24$ ) for preseasonal and SMD  $-0.86$  (95% CI  $-1.49, -0.22$ ) for continuous (results from one study only) (Figures S26A and B), these analyses showing a clear benefit from pre-/coseasonal treatment and the suggestion (but not confirming) benefit from continuous treatment
- Modified allergen extracts (allergoids) vs unmodified allergen extracts in SCIT: SMD  $-0.49$  (95% CI  $-0.79, -0.19$ ) for allergoids and SMD  $-0.36$  (95% CI  $-0.73, 0.03$ ) (Figures S27A and B), these finding a clear benefit from allergoids and suggesting (but not confirming) a benefit from unmodified preparations.

- Different allergens for AIT (SCIT and SLIT): Grass: SMD  $-0.41$  (95% CI  $-0.58, -0.24$ ) vs Tree (one study only): SMD  $-0.26$  (95% CI  $-0.64, 0.13$ ) vs Molds: SMD  $-0.65$  (95% CI  $-2.06, 0.76$ ) vs Weeds: SMD  $-0.69$  (95% CI  $-1.24, -0.13$ ) (Figure S28A-D), this showing clear evidence of benefit for grass and tree pollens, and suggesting (but not confirming) evidence of benefit for molds and weeds.

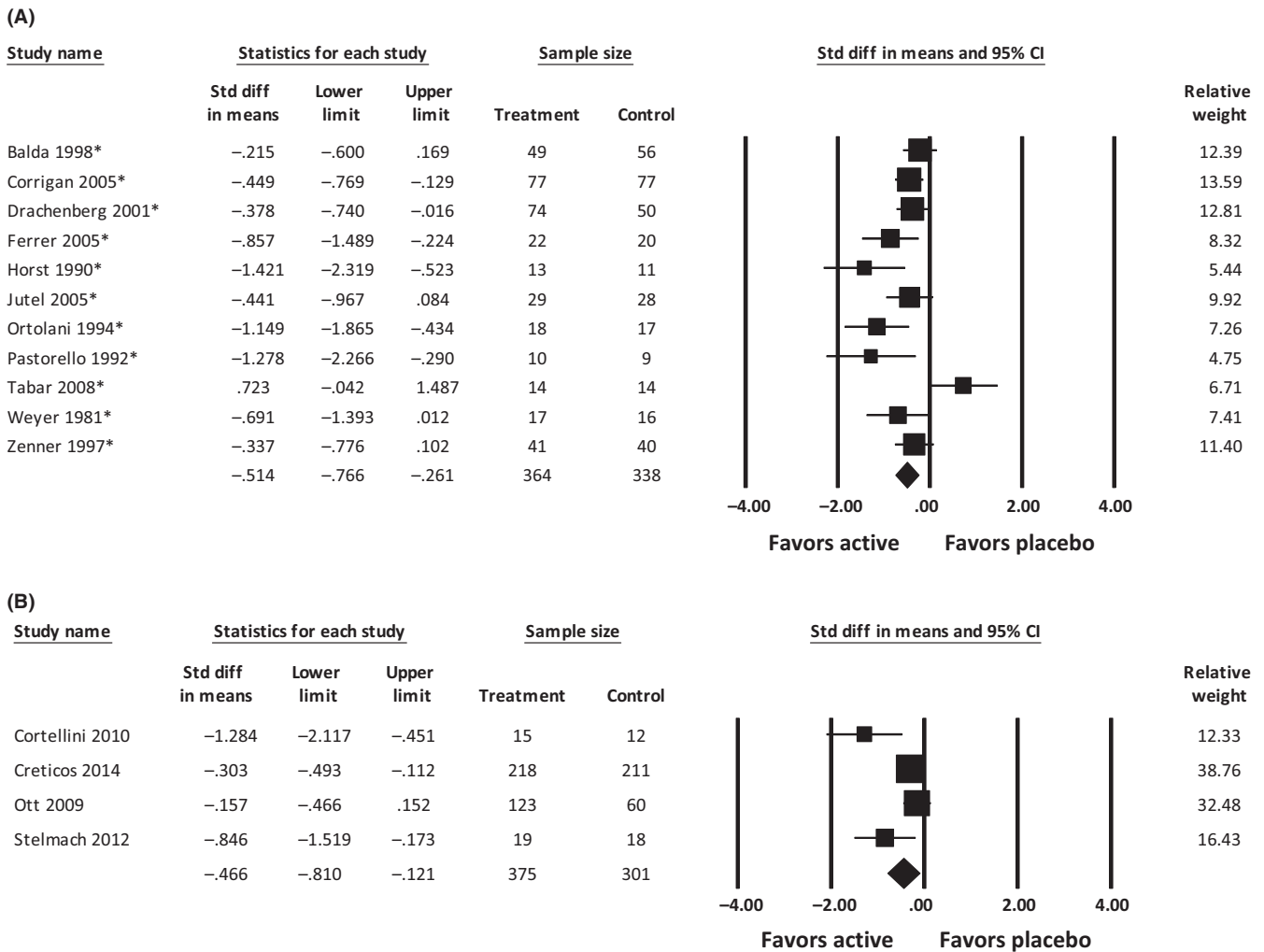
### Long-term

We found one SCIT trial<sup>53</sup> and two SLIT trials<sup>109,133</sup> that reported on this outcome. These are described in detail in the supplement. Overall, one of the three trials found evidence of a sustained beneficial effect on combined symptom and medication scores. The one trial at an unclear ROB demonstrated a two-year carryover effect of AIT in the active SLIT group that received AIT four months preseasonally for three consecutive seasons but not for the group which received AIT two months preseasonally.<sup>109,159</sup>

## 3.3 | Secondary outcomes

### 3.3.1 | Disease-specific quality of life

Thirty studies reported data on quality of life (QoL): these comprised SCIT ( $n=17$ )<sup>19,20,23,28,33-35,45,46,55,58,68-70,72,74,79</sup> and SLIT ( $n=13$ )<sup>90,99,104,106,108,110,117,129,130,132,140,145,149</sup> trials (Table S1j and k). The majority of trials ( $n=29$ ) used one of the disease-specific, validated Rhinitis Quality of Life Questionnaire (RQLQ) instruments. However, one SLIT study (eligible because it reported on other outcomes) used a generic, non-disease-specific tool, the SF-



**FIGURE 8** Meta-analysis of double-blind RCTs comparing combined symptom and medication scores between (A) subcutaneous immunotherapy (SCIT) and placebo groups and (B) sublingual immunotherapy (SLIT) and placebo groups (random-effects models). (A) Heterogeneity:  $\tau^2=.096$ ;  $\chi^2=23.777$ ,  $df=10$  ( $P<.008$ );  $I^2=58\%$ . Test for overall effect:  $Z=-3.984$  ( $P<.0001$ ). \*denotes SCIT studies. (B) Heterogeneity:  $\tau^2=.070$ ;  $\chi^2=8.584$ ,  $df=3$  ( $P<.035$ );  $I^2=65\%$ . Test for overall effect:  $Z=-2.648$  ( $P<.008$ )

36, and this was therefore not considered further.<sup>140</sup> Due to inconsistencies of reporting data, it was not possible to pool results from all of the studies and no SLIT studies were suitable for inclusion in meta-analysis. Pooling data from the six SCIT studies with suitably reported data derived from the original and standardized RQLQ instruments found a SMD of  $-0.35$  (95% CI  $-0.74, 0.04$ ), this corresponding to a likely small-to-medium improvement in the AIT group when compared to placebo (Figure 9).

### 3.3.2 | Allergen challenge models in AIT

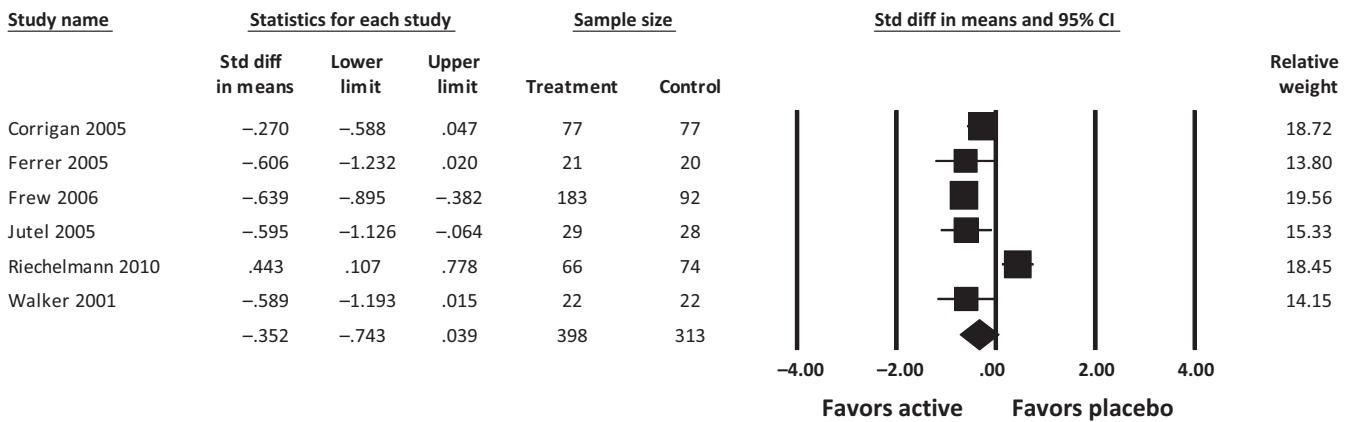
A detailed description of environmental exposure chamber, nasal and conjunctival challenge studies is described in the supplement. One SCIT and three SLIT<sup>83,120,121</sup> chamber studies demonstrated the effectiveness of AIT. Results of nasal challenge studies for 15 SCIT<sup>23,24,27,29,30,33,37,43,52,57-59,63,64,75</sup> and 11

SLIT<sup>84,86,87,92,93,122,128,136,139,146,150</sup> (Table S1I) were conflicting making it difficult to make clear conclusions. There was no clear evidence of effectiveness in 12 SCIT<sup>21,23,35,38,42,45,55,62-64,70,72</sup> and four SLIT conjunctival challenges studies<sup>120,127,138,146</sup> (Table S1m).

### 3.3.3 | Cost-effectiveness

#### Characteristics of studies

We identified 19 eligible studies that reported on health economic evaluations of SCIT and SLIT in both children and adults (Table S1n).<sup>160-178</sup> Studies were based in a range of countries. Seven of the studies reported results against disease-specific outcome measures while the remaining 12 reported results based on quality-adjusted life years (QALYs). Thirteen of the studies were based on RCT data or meta-analyses of RCT data.<sup>160-169,176-178</sup> Full details are in the supplement.



**FIGURE 9** Meta-analysis of double-blind RCTs comparing quality of life scores between subcutaneous immunotherapy (SCIT) and placebo groups (random-effects models). Heterogeneity:  $\tau^2=.186$ ;  $\chi^2=28.432$ ,  $df=5$  ( $P<.0001$ );  $I^2=82\%$ . Test for overall effect:  $Z=-1.764$  ( $P<.078$ )

### Quality appraisal

The quality appraisal of the included studies is detailed in Table S10.

### Main findings

In general, the studies found that AIT, and where defined both SLIT and SCIT, were more effective than standard care including pharmacotherapy, but also more expensive. The studies that compared SLIT with SCIT gave very mixed results not allowing a clear conclusion to be drawn that either treatment was necessarily more effective or more costly than the other from a health system perspective. The studies comparing Grazax (SLIT) and Oralair (SLIT) suggested that Oralair is both more effective and cheaper than Grazax.<sup>165,167</sup>

For those studies based on RCT data conducted from a health system perspective and using QALYs as their outcome measure ( $n=7$ ), we found that:

- Nasser 2008: In patients with both rhinitis and asthma in England the incremental cost-effectiveness ratio (ICER) for SLIT vs standard care was £8816 (€10 851) per QALY at 2005 prices inflated using national health service (NHS) inflation indices (ie, Personal Social Services Research Unit (PSSRU)) to £10 726 (€13 202) per QALY at 2014/15 prices.<sup>177</sup>
- Poulsen 2008: In adult patients with rhinoconjunctivitis in Denmark the ICER for SLIT vs standard care was 134 105 DKK per QALY (no price year was given so we assumed study year of 2008) updating to current prices and £ at 0.1 £ per DKK gave an ICER of £15 294 (€18 824) per QALY at 2014/15 prices.<sup>164</sup>
- Keiding 2007: In a study in adult patients with rhino-conjunctivitis performed in the U.K. ICERs of SCIT were calculated using health care data from Austria, Denmark, Finland, Germany, Netherlands, Sweden. The ICERs of SCIT compared to standard care in 2005 Euro per QALY were 9716, 2586, 13683, 10300, 24519 and 22675, respectively. Updating to current prices and £ at 0.75 GBP per Euro gives ICERs of £8866, £2360, £12486, £9399,

£22374 and £20691 per QALY respectively at 2014/15 prices.<sup>162</sup>

- Ronaldson 2014: In 5- to 16-year-olds with rhinoconjunctivitis with or without asthma in the UK, the ICER for SLIT vs standard care was £12 168 (€14 976) per QALY at 2008 prices. Updating to current prices gives an ICER of £13 357 (€16 440) per QALY at 2014/15 prices.<sup>166</sup>
- Westerhout 2012: In patients with rhinoconjunctivitis without asthma in Germany the ICER for SLIT (Oralair) vs standard care was 14 728 euros per QALY at 2011 prices. Converting to current prices and GBP at 0.75 £ per Euro gives an ICER of £11 460 per QALY.<sup>167</sup>
- Verheggen 2015: In patients with rhinoconjunctivitis without asthma in Germany the ICER for SLIT (Oralair) vs SCIT is 12 593 euros per QALY at 2013 prices. Converting to 2014/15 prices and GBP at 0.75 GBP per Euro gives an ICER of £9627 per QALY.<sup>168</sup>
- Reinhold 2016: In patients with rhinoconjunctivitis without asthma in Germany SCIT (Allergovit) is cheaper and more effective than SLIT (Oralair). The ICER for SCIT (Allergovit) standard care is 11 000 euros per QALY at 2013 prices. Converting to 2014/15 prices and GBP at 0.75 GBP per Euro gives an ICER of £8334 per QALY.<sup>169</sup>

When assessing these results, it was unclear how comparable the patient populations were between the studies; a key factor that impacts the costs and quality of life observed is the proportion of patients who have asthma as well as rhinitis—these proportions were not reported in the studies. Also noteworthy was that the ICERs for AIT seemed to vary substantially between different health systems as demonstrated in Keiding et al.<sup>162</sup> 2007 where ICERs range from £2360 per QALY in Denmark to £22 374 per QALY in the Netherlands suggesting that straightforward conclusions may not be generalizable even across seemingly similar countries.<sup>162</sup>

### Overall interpretation

The seven key studies identified, disregarding the caveats about generalizability, suggested that SLIT and SCIT treatment would be considered cost-effective in this patient population in England at the standard NICE cost-effectiveness threshold of £20 000 (€24 616) per QALY. However, the quality of the studies and the general lack of attention to characterizing uncertainty and handling missing data need to be taken into account when interpreting these results.<sup>162,164,166-169,177</sup>

### 3.3.4 | Safety

RCTs and case series were eligible for inclusion to consider the safety of AIT.

#### Randomized controlled trials

Safety data for SCIT and SLIT RCTs are summarized in Table S1p-v. There was a great variation in reporting of adverse events (AEs) and a number of grading scales including WAO and EAACI were used. As detailed in the tables some studies reported limited or unclear data on number of AEs, some studies reported no data on AEs and others reported that no AEs occurred at all through the duration of the trial period. Conversely some studies reported all treatment emergent AEs.

**Total AEs** We were able to pool data for this outcome for total number of AEs. Safety data for 51 SCIT and SLIT RCTs were pooled to give an overall risk ratio (RR) of experiencing an AE of 1.64 (95% CI 1.43, 1.89) (Figure S3A).

For SCIT studies (n=19), we found an RR of 1.58 (95% CI 1.13, 2.20) of experiencing an AE and for SLIT studies (n=32) an RR of 1.68 (95% CI 1.44, 1.98), (Figure S3B,C) suggesting a comparable safety profile for both modes of AIT.

**Systemic AEs** We were able to pool data for number of systemic AEs for 39 SCIT and SLIT RCTs to give an overall RR of experiencing a systemic AE of 1.26 (95% CI 1.03, 1.55) (Figure S3D). For SCIT studies (n=15), we found a RR of 1.15 (95% CI 0.67, 2.00) of experiencing a systemic AE and for SLIT studies (n=24) a RR of 1.31 (95% CI 1.05, 1.63) (Figure S3E,F).

We were able to pool data for the number of patients experiencing a systemic AE for SCIT and SLIT RCTs (n=18) to give a RR of 2.37 (95% CI 1.09, 5.16) (Figure S3G).

**Local AEs** We were able to pool data for local AEs for 39 SCIT and SLIT RCTs to give an overall RR of experiencing a local AE of 1.78 (95% CI 1.51, 2.11) (Figure S3H). For SCIT studies (n=9), we found an RR of 2.21 (95% CI 1.43, 3.41) of experiencing a local AE and for SLIT studies (n=30) an RR of 1.71(95% CI 1.43, 2.05) (Figure S3I,J).

We were able to pool data for the number of patients experiencing a local AE for SCIT and SLIT RCTs (n=17) to give a RR of 1.72 (95% CI 1.32, 2.23) (Figure S3K).

### Case series

Seven large case series were identified.<sup>179-185</sup> (Table S1w-y) Local (LR) and systemic (SR) AEs were recorded in a range of treatment protocols, including conventional, rush, ultrarush and cluster. In total 4045 patients were included in these case series however only 3541 were patients with allergic rhinoconjunctivitis; we therefore focused on data for these patients.

The case series were conducted in a number of countries including Spain, Colombia, the USA, Germany and Portugal.

The case series highlighted that where modified allergen extracts were used to deliver AIT this was safer in terms of number of AEs reported compared to unmodified extracts.<sup>180-183</sup>

Safety data from the rush<sup>180</sup> and ultrarush<sup>181,182</sup> protocols were evaluated and are presented in Table S1 w and x. The studies concluded that the frequency of SRs were similar to conventional buildup schedules, but importantly rush and ultrarush protocols were associated with improved patient adherence to treatment by reducing the number of injections required and the cost associated with treatment. Comparable benefits of cluster treatment protocol were also reported in one study.<sup>184</sup> Finally, one case series looked at investigating the number of AEs where patients received either conventional or cluster IT via the SLIT route. AEs were reported in 0.15% of all administered doses in which 9.3% of patients experienced a SR. The study concluded that SLIT was safe in the treatment of allergic rhinoconjunctivitis.<sup>179</sup>

No fatalities were reported in any of these studies.

## 4 | DISCUSSION

### 4.1 | Statement of principal findings

This review of a very substantial body of international trial evidence, many of which were judged to be at low ROB, has found clear evidence that AIT improved all three of our primary outcomes—that is, symptom, medication, and combined symptom and medication scores over the short term. These findings were robust to prespecified sensitivity analyses but evidence of potential publication bias was identified for all three primary outcomes. Although the long-term studies are fewer in number, there was a modest evidence-base in support of the effectiveness of AIT in improving symptom scores after treatment discontinuation for both SCIT and SLIT. The evidence was less clear in relation to the impact on medication and combined symptom and medication scores. SCIT improved disease-specific quality of life. We could draw no clear conclusions on the effectiveness of AIT on nasal and conjunctival challenges and on cost-effectiveness which may be cost-effective in an English NHS setting, but due to the poor quality of the studies this needs to be interpreted with caution. AIT increased the risk of AEs for both SCIT and SLIT, but no fatalities occurred.

### 4.2 | Strengths and limitations

To our knowledge, this is the most comprehensive assessment of AIT in allergic rhinoconjunctivitis ever undertaken. We employed internationally accepted techniques to systematically identify, assess, and synthesize a substantial body of evidence. This involved taking



advantage of and building on other recent systematic reviews focusing on distinct modes of delivering AIT.

The limitations of this review need to be considered. First, despite our extensive searches we may not have uncovered all relevant evidence on this subject. Second, we were limited by the heterogeneity in approaches used to assess outcomes, which meant we were unable to pool data from all trials or undertake all the planned subgroup analyses. Furthermore studies for which data was pooled also showed heterogeneity which may be related to the diverse populations studied, protocols followed, products used and duration of trial period. For the subgroup analyses that were undertaken, there was in some cases imprecision which impacted on our ability to draw clear conclusions. These subgroup analyses were indirect comparisons between SCIT and SLIT and the findings should therefore be cautiously interpreted. Third, because of the heterogeneity in scoring systems used, we undertook meta-analyses using random-effects modeling and pooled data using SMDs, which can be difficult to interpret. The absolute size of the SMD was used to guide assessment of the likely effect size demonstrated.<sup>186</sup> Finally, it needs to be borne in mind that there may have been important differences in effectiveness between specific AIT products. Investigating this issue was however beyond the scope of this review. In terms of safety there was heterogeneity in reporting of AEs with many differing scoring systems used due to this we were unable to report this outcome as originally planned using only the WAO grading system.

### 4.3 | Implications for policy, practice, and research

Our findings clearly show that AIT is effective in improving the three patient-reported outcomes that represented our primary outcomes, at least over the short term, and that AIT should therefore be considered in the management of patients with allergic rhinoconjunctivitis.

Greater standardization of trial designs and reporting techniques—in particular, in relation to choice of outcomes and their reporting so as to facilitate evidence syntheses and key subgroup analyses, would greatly help to advance the research base underpinning AIT. We therefore appreciate initiatives of the EAACI in, for example, harmonizing and standardizing clinical endpoints in AIT<sup>187</sup> or determining threshold level of relevant pollen seasons for assessing clinical effect sizes.<sup>188</sup> We also wish to highlight the need for additional studies focusing on long-term outcomes and on studies of ILIT and other novel modes of delivery. We hope that future researchers will build on the findings from this systematic review and aim to fill key evidence gaps and areas of continuing uncertainty.

The findings from this review will be used to inform the development of recommendations for EAACI's Guidelines on AIT for Allergic Rhinoconjunctivitis.

## 5 | CONCLUSIONS

AIT is effective in achieving clinically important short-term improvements in symptom, medication, and combined symptom and

medication scores. There is a limited body of evidence on the longer-term effectiveness of AIT in improving symptom scores.

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### CONFLICT OF INTERESTS

S. Dhami reports grants from EAACI to carry out the review, during the conduct of the study. U. Nurmatov, S. Arasi, and M. Asaria report payment from Evidence-Based Health Care Ltd during the conduct of the study. G. Roberts has a patent issued: "Use of sublingual immunotherapy to prevent the development of allergy in at risk infants"; and his university has received payments for the activities he has undertaken: giving expert advice to ALK, and presenting at company symposia for ALK, Allergen Therapeutics, and Meda, and serving as a member of an Independent Data Monitoring Committee for Merck. O. Pfaar reports grants and personal fees from ALK-Abelló, Allergopharma, Stallergenes Greer, HAL-Allergy Holding B.V./HAL-Allergie GmbH, Bencard Allergie GmbH/Allergy Therapeutics, Lofarma, Biotech Tools S.A., Laboratorios LETI/LETI Pharma, and Anergis S.A.; grants from Biomay, Nuvo, and Circassia; and personal fees from MEDA Pharma, Sanofi US Services, Mobile Chamber Experts (a GA<sup>2</sup>LEN Partner), Novartis Pharma and Pohl-Boskamp, outside the submitted work. A. Muraro reports personal fees from Novartis, Meda, and Mylan, outside the submitted work. IJ. Ansoategui reports personal fees from SANOFI, Bayer, Pfizer, FAES FARMA, MIT FARMA, HIKMA, Menarini, and Bial Aristegui, outside the submitted work. M. Calderon has received honorarium in advisory boards for ALK and Hal-Allergy and served as a speaker for ALK, Merck, and Stallergenes Greer. S. Durham reports grants from Regeneron (USA), Biotech Tools, ALK (Denmark), Food Standards Agency (UK), and National Institute of Health Research (UK) and personal fees from Anergis (Switzerland), Circassia (UK), Biomay (Austria), Merck, Allergy Therapeutics (UK), ALK (Hørsholm, Denmark), med update GmbH (Germany), and Allergy Therapeutics, outside the submitted work. R. Gerth van Wijk reports personal fees from ALK-Abelló, Circassia, and Allergopharma, during the conduct of the study. S. Halken reports personal fees from ALK-Abelló and from different companies, for example, Meda, Stallergenes, Allergopharma, and ALK-Abelló, outside the submitted work. E. Hamelmann is giving lectures in industry symposia and takes part in advisory board meetings for the following companies: Boehringer Ingelheim, Novartis, ALK, HAL-Allergy, Bencard, Stallergenes, Leti Pharma, and SymbioPharm. L. Jacobsen reports personal fees from EAMG, outside the submitted work. D. Larenas-Linnemann has no influence in the presented paper. R. Mosges reports personal fees from ALK, Allergopharma, Allergy Therapeutics, Friulchem, Hexal, Servier, Klosterfrau, Bayer, FAES, GSK, MSD, Johnson&Johnson,



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## AUTHOR CONTRIBUTIONS

This review was drafted by S. Dhami, U. Nurmatov, and A. Sheikh. It was initially revised following critical review by G. Roberts and O. Pfaar and then by all co-authors. This paper is part of the EAACI AIT guidelines project, chaired by Antonella Muraro and coordinated by Graham Roberts.

## REFERENCES

- Mallol J, Crane J, von Mutius E, Odhiambo J, Keil U, Stewart A. International study of asthma and allergies in childhood (ISAAC) phase three: a global synthesis. *Allergol Immunopathol (Madr)*. 2013;41:73-85.
- Patil VK, Kurukulaaratchy RJ, Venter C, et al. Changing prevalence of wheeze, rhinitis and allergic sensitisation in late childhood: findings from 2 Isle of Wight birth cohorts 12 years apart. *Clin Exp Allergy*. 2015;45:1430-1438.
- Bousquet J, Khaltaev N, Cruz AA, et al. Allergic rhinitis and its impact on asthma (ARIA) 2008 update (in collaboration with the World Health Organization, GA(2)LEN and AllerGen. *Allergy*. 2008;63(Suppl 86):8-160.
- Skoner DP. Allergic rhinitis: definition, epidemiology, pathophysiology, detection, and diagnosis. *J Allergy Clin Immunol*. 2001;108(1 Suppl):S2-S8.
- Walker S, Khan-Wasti S, Fletcher M, Cullinan P, Harris J, Sheikh A. Seasonal allergic rhinitis is associated with a detrimental effect on examination performance in United Kingdom teenagers: case-control study. *J Allergy Clin Immunol*. 2007;120:381-387.
- Blanc PD, Trupin L, Eisner M, et al. The work impact of asthma and rhinitis: findings from a population-based survey. *J Clin Epidemiol*. 2001;54:610-618.

- Guerra S1, Sherrill DL, Martinez FD, Barbee RA. Rhinitis as an independent risk factor for adult-onset asthma. *J Allergy Clin Immunol*. 2002;109:419-425.
- Zuberbier T, Bachert C, Bousquet PJ, et al. GA<sup>2</sup>LEN/EAACI pocket guide for allergen-specific immunotherapy for allergic rhinitis and asthma. *Allergy*. 2010;65:1525-1530.
- Walker SM, Durham SR, Till SJ, et al. Immunotherapy for allergic rhinitis. *Clin Exp Allergy*. 2011;41:1177-1200.
- Pfaar O, Bachert C, Bufe A, et al. Guideline on allergen-specific immunotherapy in IgE mediated allergic diseases – S2k Guideline of the German Society for Allergology and Clinical Immunology (DGAKI), the Society for Pediatric Allergy and Environmental Medicine (GPA), the Medical Association of German Allergologists (AeDA), the Austrian Society for Allergy and Immunology (ÖGAI), the Swiss Society for Allergy and Immunology (SGAI), the German Society of Dermatology (DDG), the German Society of Oto-Rhino-Laryngology, Head and Neck Surgery (DGHNO-KHC), the German Society of Pediatrics and Adolescent Medicine (DGKJ), the Society for Pediatric Pneumology (GPP), the German Respiratory Society (DGP), the German Association of ENT Surgeons (BV-HNO), the Professional Federation of Paediatricians and Youth Doctors (BVKJ), the Federal Association of Pulmonologists (BDP) and the German Dermatologists Association (BVDD). *Allergo J Int*. 2014; 23:282-319.
- Hellings PW, Fokkens WJ, Akdis C, et al. Uncontrolled allergic rhinitis and chronic rhinosinusitis: where do we stand today? *Allergy*. 2013;68:1-7.
- Dhami S, Nurmatov U, Roberts G, et al. Allergen immunotherapy for allergic rhinoconjunctivitis: protocol for a systematic review. *Clin Transl Allergy*. 2016;6:12.
- Cochrane Risk of bias tool. [http://handbook.cochrane.org/chapter\\_8/table\\_8\\_5\\_a\\_the\\_cochrane\\_collaborations\\_tool\\_for\\_assessing.htm](http://handbook.cochrane.org/chapter_8/table_8_5_a_the_cochrane_collaborations_tool_for_assessing.htm) Accessed 3 January 2017
- CASP checklist for Economic evaluations. [http://media.wix.com/ugd/dded87\\_3b2bd5743feb4b1aaac6ebdd68771d3f.pdf](http://media.wix.com/ugd/dded87_3b2bd5743feb4b1aaac6ebdd68771d3f.pdf). Accessed 3 January 2017.
- NICE Case Series Risk of Bias tool. <https://www.nice.org.uk/guidance/cg3/resources/appendix-4-quality-of-case-series-form2>. Accessed 3 January 2017.
- Higgins JPT, Green S. *Cochrane Handbook for Systematic Reviews of Interventions*. Version 5.0.2 (Chapter 11, Section 11). The Cochrane Collaboration, 2011. Available from <http://handbook.cochrane.org>.
- Egger M, Davey Smith G, Schneider M, et al. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315:34.
- Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics*. 1988;44:101-110.
- Alvarez-Cuesta E, Aragonese-Gilsanz E, Martin-Garcia C, Berges-Gimeno P, Gonzalez-Mancebo E, Cuesta-Herranz J. Immunotherapy with depigmented glutaraldehyde-polymerized extracts: changes in quality of life.[Erratum appears in Clin Exp Allergy. 2005 Nov; 35 (11):1504]. *Clin Exp Allergy*. 2005;35:572-578.
- Ariano R, Kroon AM, Augeri G, Canonica W, Passalacqua G. Long-term treatment with allergoid immunotherapy with Parietaria. Clinical and immunologic effects in a randomized, controlled trial. *Allergy*. 1995;54:313-319.
- Arvidsson MB, Lowhagen O, Rak S. Effect of 2-year placebo-controlled immunotherapy on airway symptoms and medication in patients with birch pollen allergy. *J Allergy Clin Immunol*. 2002;109:777-783.
- Balda BR, Wolf H, Baumgarten C, et al. Tree-pollen allergy is efficiently treated by short-term immunotherapy (STI) with seven pre-seasonal injections of molecular standardized allergens. *Allergy*. 1998;53:740-748.
- Bodtger U, Poulsen LK, Jacobi HH, Malling HJ. The safety and efficacy of subcutaneous birch pollen immunotherapy – a one-year,

- randomised, double-blind, placebo-controlled study. *Allergy*. 2002;57:297-305.
24. Bousquet J, Hejjaoui A, Skassa-Brociek W, et al. Double-blind, placebo-controlled immunotherapy with mixed grass-pollen allergoids. I. Rush immunotherapy with allergoids and standardized orchard grass-pollen extract. *J Allergy Clin Immunol*. 1987;80:591-598.
  25. Bousquet J, Hejjaoui A, Soussana M, et al. Double-blind, placebo-controlled immunotherapy with mixed grass-pollen allergoids IV. Comparison of the safety and efficacy of two dosages of a high-molecular-weight allergoid. *J Allergy Clin Immunol*. 1990;85:490-497.
  26. Bousquet J, Maasch HJ, Hejjaoui SA, et al. Double-blind, placebo-controlled immunotherapy with mixed grass-pollen allergoids III. Efficacy and safety of unfractionated and high-molecular-weight preparations in rhinoconjunctivitis and asthma. *J Allergy Clinical Immunol*. 1989;84:546-556.
  27. Bousquet J, Becker WM, Hejjaoui A, et al. Differences in clinical and immunologic reactivity of patients allergic to grass pollens and to multiple-pollen species II. Efficacy of a double-blind, placebo-controlled, specific immunotherapy with standardized extracts. *J Allergy Clinical Immunol*. 1991;88:43-53.
  28. Bozek A, Kolodziejczyk K, Krajewska-Wojtys A, Jarzab J. Pre-seasonal, subcutaneous immunotherapy: a double-blinded, placebo-controlled study in elderly patients with an allergy to grass. *Ann Allergy Asthma Immunol*. 2016;116:156-161.
  29. Brunet C, Bedard PM, Lavoie A, et al. Allergic rhinitis to ragweed pollen I. Reassessment of the effects of immunotherapy cellular and humoral responses. *J Allergy Clinical Immunol*. 1992;89:76-86.
  30. Horst M, Hejjaoui A, Horst V, Michel FB, Bousquet J. Double-blind, placebo-controlled rush immunotherapy with a standardized Alternaria extract. *J Allergy Clin Immunol*. 1990;85:460-472.
  31. Ceuppens JL, Bullens D, Kleinjans H, van der Werf J, Purethal Birch Efficacy Study Group. Immunotherapy with a modified birch pollen extract in allergic rhinoconjunctivitis: clinical and immunological effects. [Erratum appears in *Clin Exp Allergy*. 2012 Oct;42(10):1543]. *Clin Exp Allergy*. 2009;39:1903-1909.
  32. Chakraborty P, Roy I, Chatterjee S, Chanda S, Gupta-Bharracharya S. Phoenix sylvestris Roxb pollen allergy: a 2-year randomized controlled trial and follow-up study of immunotherapy in patients with seasonal allergy in an agricultural area of West Bengal, India. *J Invest Allergol Clin Immunol*. 2006;16:377-384.
  33. Charpin D, Gouitaa M, Dron-Gonzalez M, et al. Immunotherapy with an aluminum hydroxide-adsorbed Juniperus ashei foreign pollen extract in seasonal indigenous cypress pollen rhinoconjunctivitis. A double-blind, placebo-controlled study. *Int Arch Allergy Immunol*. 2007;143:83-91.
  34. Colas C, Monzon S, Venturini M, Lezaun A. Double-blind, placebo-controlled study with a modified therapeutic vaccine of Salsola kali (Russian thistle) administered through use of a cluster schedule. *J Allergy Clin Immunol*. 2006;117:810-816.
  35. Corrigan CJ, Kettner J, Doemer C, Cromwell O, Narkus A, for the Study Group\*. Efficacy and safety of preseasonal-specific immunotherapy with an aluminium-adsorbed six-grass pollen allergoid. *Allergy*. 2005;60:801-807.
  36. Crimi N, Li Gotti F, Mangano G, et al. A randomized, controlled study of specific immunotherapy in monosensitized subjects with seasonal rhinitis: effect on bronchial hyperresponsiveness, sputum inflammatory markers and development of asthma symptoms. *Ann Ital Med Int*. 2004;19:98-108.
  37. Dokic D, Schnitker J, Narkus A, Cromwell O, Frank E. Clinical effects of specific immunotherapy: a two-year double-blind, placebo-controlled study with a one year follow-up. *Prilozi*. 2005;26:113-129.
  38. Dolz I, Martinez-Cocera C, Bartolome JM, Cimarra M. A double-blind, placebo-controlled study of immunotherapy with grass-pollen extract Alutard SQ during a 3-year period with initial rush immunotherapy. *Allergy*. 1996;51:489-500.
  39. Drachenberg KJ, Wheeler AW, Stuebner P, Horak F. A well-tolerated grass pollen-specific allergy vaccine containing a novel adjuvant, monophosphoryl lipid A, reduces allergic symptoms after only four preseasonal injections. *Allergy*. 2001;56:498-505.
  40. Drachenberg K, Heinzkill M, Urban E. Short-term immunotherapy with tree pollen allergoids and the adjuvant monophosphoryl lipid-A – Results from a multicentre, placebo-controlled, randomised, double-blind study. [Kurzeit-Immuntherapie mit Baumpollen-Allergoiden und dem Adjuvans Monophosphoryl Lipid-A]. *Allergologie*. 2002;25:466-474.
  41. DuBuske L, Frew A, Horak F, Keith P, Corrigan C, Aberer W. Ultra-short-specific immunotherapy successfully treats seasonal allergic rhinoconjunctivitis to grass pollen. *Allergy Asthma Proc*. 2011;32:239-247.
  42. Durham SR, Walker SM, Varga EM, et al. Long-term clinical efficacy of grass-pollen immunotherapy. *N Engl J Med*. 1999;341:468-475.
  43. Ewan PW, Alexander MM, Snape C, Ind PW, Agrell B, Dreborg S. Effective hyposensitization in allergic rhinitis using a potent partially purified extract of house dust mite. *Clin Allergy*. 1988;18:501-508.
  44. Fell P, Brostoff J. A single dose desensitization for summer hay fever. Results of a double blind study-1988. *Eur J Clin Pharmacol*. 1990;38:77-79.
  45. Ferrer M, Burches E, Peláez A, et al. Double-blind, placebo-controlled study of immunotherapy with Parietaria judaica: clinical efficacy and tolerance. *J Invest Allergol Clin Immunol*. 2005;15:283-292.
  46. Frew AJ, Powell RJ, Corrigan CJ, Durham SR, UK Immunotherapy Study Group. Efficacy and safety of specific immunotherapy with SQ allergen extract in treatment-resistant seasonal allergic rhinoconjunctivitis. *J Allergy Clin Immunol*. 2006;117:319-325.
  47. Grammer LC, Zeiss CR, Suszko IM, et al. A double-blind, placebo-controlled trial of polymerized whole ragweed for immunotherapy of ragweed allergy. *Allergy Clin Immunology*. 1982;6:494-499.
  48. Grammer LC, Shaughnessy MA, Suszko IM, Shaughnessy JJ, Patterson R. A double-blind histamine placebo-controlled trial of polymerized whole grass for immunotherapy of grass allergy. *J Allergy Clin Immunol*. 1983;72:448-453.
  49. Grammer LC, Shaughnessy MA, Suszko IM, et al. Persistence of efficacy after a brief course of polymerized ragweed allergen: a controlled study. *J Allergy Clin Immunol*. 1984;73:484-489.
  50. Grammer LC, Shaughnessy MA, Bernhard MI, et al. The safety and activity of polymerized ragweed: a double-blind, placebo-controlled trial in 81 patients with ragweed rhinitis. *J Allergy Clin Immunol*. 1987;80:177-183.
  51. Hoiby AS, Strand V, Robinson DS, Sager A, Rak S. Efficacy, safety, and immunological effects of a 2-year immunotherapy with Depigoid birch pollen extract: a randomized, double-blind, placebo-controlled study. *Clin Exp Allergy*. 2010;40:1062-1070.
  52. Iliopoulos O, Proud D, Franklin Adkinson Jr N, et al. Effects of immunotherapy on the early, late, and rechallenge nasal reaction to provocation with allergen: changes in inflammatory mediators and cells. *J Allergy Clin Immunol*. 1991;4:855-866.
  53. James LK, Shamji MH, Walker SM, et al. Long-term tolerance after allergen immunotherapy is accompanied by selective persistence of blocking antibodies. *J Allergy Clin Immunol*. 2011;127:509-516.
  54. Juniper EF, Kline PA, Ramsdale EH, Hargreave FE. Comparison of the efficacy and side effects of aqueous steroid nasal spray (budesonide) and allergen-injection therapy (Pollinex-R) in the treatment of seasonal allergic rhinoconjunctivitis. *J Allergy Clin Immunol*. 1990;85:606-611.
  55. Jutel M, Jaeger L, Suck R, Meyer H, Fiebig H, Cromwell O. Allergen-specific immunotherapy with recombinant grass pollen allergens. *J Allergy Clin Immunol*. 2005;116:608-613.

56. Kleine-Tebbe J, Walmar M, Bitsch-Jensen K, et al. Negative clinical results from a randomised, double-blind, placebo-controlled trial evaluating the efficacy of two doses of immunologically enhanced, grass subcutaneous immunotherapy despite dose-dependent immunological response. *Clin Drug Investig.* 2014;34:577-586.
57. Klimek L, Uhlig J, Mosges R, Rettig K, Pfaar O. A high polymerized grass pollen extract is efficacious and safe in a randomized double-blind, placebo-controlled study using a novel up-dosing cluster-protocol. *Allergy.* 2014;69:1629-1638.
58. Kuna P, Kaczmarek J, Kupczyk M. Efficacy and safety of immunotherapy for allergies to *Alternaria alternata* in children. *J Allergy Clin Immunol.* 2011;127:502-508.
59. Leynadier F, Banoun L, Dollois B, et al. Immunotherapy with a calcium phosphate-adsorbed five-grass-pollen extract in seasonal rhinoconjunctivitis: a double-blind, placebo-controlled study. *Clin Exp Allergy.* 2001;31:988-996.
60. Metzger WJ, Dorminey HC, Richerson HB, et al. Clinical and immunologic evaluation of glutaraldehyde-modified, tyrosine-adsorbed short ragweed extract: a double-blind, placebo-controlled trial. *J Allergy Clin Immunol.* 1981;68:442-448.
61. Mirone C, Albert F, Tosi A, et al. Efficacy and safety of subcutaneous immunotherapy with a biologically standardized extract of *Ambrosia artemisiifolia* pollen: a double-blind, placebo-controlled study. *Clin Exp Allergy.* 2004;34:1408-1414.
62. Olsen OT, Frølund L, Heinig J, Jacobsen L, Svendsen UG. A double-blind, randomized study investigating the efficacy and specificity of immunotherapy with *Artemisia vulgaris* or *Phleum pratense/betula verrucosa*. *Allergol Immunopathol (Madr).* 1995;23:73-78.
63. Ortolatii C, Pastorcllo EA, Iticorvaia C, et al. A double-blind, placebo-controlled study of immunotherapy with an alginate-conjugated extract of *Parietaria Judaica* in patients with *Parietaria* hay fever. *Allergy.* 1994;49:13-21.
64. Pastorello EA, Pravettoni V, Incorvaia C, et al. Clinical and immunological effects of immunotherapy with alum-adsorbed grass allergoid in grass-pollen-induced hay fever. *Allergy.* 1992;47: 281-290.
65. Patel D, Couroux P, Hickey P, et al. Fel d 1-derived peptide antigen desensitization shows a persistent treatment effect 1 year after the start of dosing: a randomized, placebo-controlled study. *J Allergy Clin Immunol.* 2012;131:103-109.
66. Pauli G, Larsen TH, Rak S, et al. Efficacy of recombinant birch pollen vaccine for the treatment of birch-allergic rhinoconjunctivitis. [Erratum appears in *J Allergy Clin Immunol.* 2009 Jan; 123 (1):166 Note: Valenta, Rudolph [corrected to Valenta, Rudolf]]. *J Allergy Clin Immunol.* 2008;122:951-960.
67. Pfaar O, Robinson D, Sager A, Emuzyte R. Immunotherapy with depigmented-polymerized mixed tree pollen extract: a clinical trial and responder analysis. *Allergy.* 2010;65:1614-1621.
68. Pfaar O, Urry Z, Robinson DS, et al. A randomized placebo-controlled trial of rush preseasonal depigmented polymerized grass pollen immunotherapy. *Allergy.* 2012;67:272-279.
69. Powell RJ, Frew AJ, Corrigan CJ, Durham SR. Effect of grass pollen immunotherapy with Alutard SQ on quality of life in seasonal allergic rhinoconjunctivitis. *Allergy.* 2007;62:1335-1338.
70. Radcliffe MJ, Lewith GT, Turner RG, Prescott P, Church MK, Holgate ST. Enzyme potentiated desensitisation in treatment of seasonal allergic rhinitis: double blind randomised controlled study. *BMJ.* 2003;327:251-254.
71. Rak S, Heinrich C, Jacobsen L, Scheynius A, Venge P. A double-blind, comparative study of the effects of short pre-season specific immunotherapy and topical steroids in patients with allergic rhinoconjunctivitis and asthma. *J Allergy Clin Immunol.* 2001;108:921-928.
72. Riechelmann H, Schmutzhard J, van der Werf JF, Distler A, Kleinjans HA. Efficacy and safety of a glutaraldehyde-modified house dust mite extract in allergic rhinitis. *Am J Rhinol Allergy.* 2010;24:104-109.
73. Tabar AI, Echechipia S, Garcia BE, et al. Double-blind comparative study of cluster and conventional immunotherapy schedules with *Dermatophagoides pteronyssinus*. *J Allergy Clin Immunol.* 2005;116: 109-118.
74. Tabar AI, Lizaso MT, Garcia BE, et al. Double-blind, placebo-controlled study of *Alternaria alternata* immunotherapy: clinical efficacy and safety. *Pediatr Allergy Immunol.* 2008;19:67-75.
75. Tari MG, Mancino M, Ghezzi E, Frank E, Cromwell O. Immunotherapy with an alum-adsorbed *Parietaria*-pollen allergoid: a 2-year, double-blind, placebo-controlled study. *Allergy.* 1997;52:65-74.
76. Tworek D, Bochenska-Marciniak M, Kuprys-Lipinska I, Kupczyk M, Kuna P. Perennial is more effective than pre-seasonal subcutaneous immunotherapy in the treatment of seasonal allergic rhinoconjunctivitis. *Am J Rhinol Allergy.* 2013;27:304-308.
77. Varney VA, Tabbah K, Mavroleon G, Frew AJ. Usefulness of specific immunotherapy in patients with severe perennial allergic rhinitis induced by house dust mite: a double-blind, randomized, placebo-controlled trial. *Clin Exp Allergy.* 1991;33:1076-1082.
78. Varney VA, Gaga M, Frew AJ, Aber VR, Kay AB, Durham SR. Usefulness of immunotherapy in patients with severe summer hay fever uncontrolled by antiallergic drugs. *BMJ.* 1991;302:265-269.
79. Walker SM, Pajno GB, Lima MT, Wilson DR, Durham SR. Grass pollen immunotherapy for seasonal rhinitis and asthma: a randomized, controlled trial. *J Allergy Clin Immunol.* 2001;107:87-93.
80. Weyer A, Donat N, L'Heritier C, et al. Grass pollen hyposensitization versus placebo therapy. I. Clinical effectiveness and methodological aspects of a pre-seasonal course of desensitization with a four-grass pollen extract. *Allergy.* 1981;36:309-317.
81. Zenner HP, Baumgarten C, Rasp G, et al. Short-term immunotherapy: a prospective, randomized, double-blind, placebo-controlled multicenter study of molecular standardized grass and rye allergens in patients with grass pollen-induced allergic rhinitis. *J Allergy Clin Immunol.* 1997;100:23-29.
82. Ahmadi-farshar A, Maarefvand M, Taymourzade B, Mazloomzadeh S, Torabi Z. Efficacy of sublingual swallow immunotherapy in children with rye grass pollen allergic rhinitis: a double-blind placebo-controlled study. *Iran J Allergy Asthma Immunol.* 2012;11:175-181.
83. Alvarez-Cuesta E, Berges-Gimeno P, Gonzalez-Mancebo E, Fernandez-Caldas E, Cuesta-Herranz J, Casanovas M. Sublingual immunotherapy with a standardized cat dander extract: evaluation of efficacy in a double blind placebo controlled study. [Erratum appears in *Allergy.* 2007 Sep; 62(9):1100 Note: Mancebo, E G [corrected to Gonzalez-Mancebo, E]]. *Allergy.* 2007;62:810-817.
84. Amar SM, Harbeck RJ, Sills M, Silveira LJ, O'Brien H, Nelson HS. Response to sublingual immunotherapy with grass pollen extract: monotherapy versus combination in a multiallergen extract. *J Allergy Clin Immunol.* 2009;124:150-156.
85. Andre C, Perrin-Fayolle M, Grosclaude M, et al. A double-blind placebo-controlled evaluation of sublingual immunotherapy with a standardized ragweed extract in patients with seasonal rhinitis. Evidence for a dose-response relationship. *Int Arch Allergy Immunol.* 2003;131:111-118.
86. Ariano R, Spadolini I, Panzani RC. Efficacy of sublingual specific immunotherapy in Cupressaceae allergy using an extract of *Cupressus arizonica*. A double blind study. *Allergol Immunopathol (Madr).* 2001;29:238-244.
87. Aydoğan M, Eifan AO, Keles S, et al. Sublingual immunotherapy in children with allergic rhinoconjunctivitis mono-sensitized to house-dust-mites: a double-blind-placebo-controlled randomised trial. *Respir Med.* 2013;107:1322-1329.
88. Bahçeciler NN, İki Uİ, Barlan İB, Başaran MM. Efficacy of sublingual immunotherapy in children with asthma and rhinitis: a double-blind, placebo-controlled study. *Pediatr Pulmonol.* 2001;32:49-55.

89. Bergmann KC, Demoly P, Worm M, et al. Efficacy and safety of sublingual tablets of house dust mite allergen extracts in adults with allergic rhinitis. *J Allergy Clin Immunol.* 2013;133:1608-1614.
90. Blaiss M, Maloney J, Nolte H, Gawchik S, Yao R, Skoner SP. Efficacy and safety of timothy grass allergy immunotherapy tablets in North American children and adolescents. *Pediatrics.* 2011;128(Suppl. 3):S136.
91. Bowen T, Greenbaum J, Charbonneau Y, et al. Canadian trial of sublingual swallow immunotherapy for ragweed rhinoconjunctivitis. *Ann Allergy Asthma Immunol.* 2004;93:425-430.
92. Bozek A, Ignasiak B, Filipowska B, Jarzab J. House dust mite sublingual immunotherapy: a double-blind, placebo-controlled study in elderly patients with allergic rhinitis. *Clin Exp Allergy.* 2012;43:242-248.
93. Bozek A, Kolodziejczyk K, Warkocka-Szoltysek B, Jarzab J. Grass pollen sublingual immunotherapy: a double-blind, placebo-controlled study in elderly patients with seasonal allergic rhinitis. *Am J Rhinol Allergy.* 2014;28:423-427.
94. Bufe A, Ziegler-Kirbach E, Stoeckmann E, et al. Efficacy of sublingual swallow immunotherapy in children with severe grass pollen allergic symptoms: a double-blind placebo-controlled study. *Allergy.* 2004;59:498-504.
95. Bufe A, Eberle P, Franke-Beckmann E, et al. Safety and efficacy in children of an SQ-standardized grass allergen tablet for sublingual immunotherapy. *J Allergy Clin Immunol.* 2009;123:167-173.
96. Caffarelli C, Sensi LG, Marcucci F, Cavagni G. Preseasonal local allergoid immunotherapy to grass pollen in children: a double-blind, placebo-controlled, randomized trial. *Allergy.* 2000;55:1142-1147.
97. Clavel R, Bousquet J, Andre C. Clinical efficacy of sublingual-swallow immunotherapy: a double-blind, placebo-controlled trial of a standardized five-grass-pollen extract in rhinitis. *Allergy.* 1998;53:493-498.
98. Cortellini G, Spadolini I, Patella V, et al. Sublingual immunotherapy for Alternaria-induced allergic rhinitis: a randomized placebo-controlled trial. *Ann Allergy Asthma Immunol.* 2010;105:382-386.
99. Cox LS, Casale TB, Nayak AS, et al. Clinical efficacy of 300IR 5-grass pollen sublingual tablet in a US study: the importance of allergen-specific serum IgE. *J Allergy Clin Immunol.* 2012;130:1327-1334.
100. Creticos PS, Maloney J, Bernstein DI, et al. Randomized controlled trial of a ragweed allergy immunotherapy tablet in North American and European adults. *J Allergy Clin Immunol.* 2013;131:1342-1349.
101. Creticos PS, Esch RE, Couroud P, et al. Randomized, double-blind, placebo-controlled trial of standardized ragweed sublingual-liquid immunotherapy for allergic rhinoconjunctivitis. [Erratum appears in *J Allergy Clin Immunol.* 2014; 133:1502]. *J Allergy Clin Immunol.* 2013;133:751-758.
102. Dahl R, Kapp A, Colombo G, et al. Efficacy and safety of sublingual immunotherapy with grass allergen tablets for seasonal allergic rhinoconjunctivitis. *J Allergy Clin Immunol.* 2006;118:434-440.
103. Dahl R, Stender A, Rak S. Specific immunotherapy with SQ standardized grass allergen tablets in asthmatics with rhinoconjunctivitis. *Allergy.* 2006;61:185-190.
104. de Blay F, Barnig C, Kanny G, et al. Sublingual-swallow immunotherapy with standardized 3-grass pollen extract: a double-blind, placebo-controlled study. *Ann Allergy Asthma Immunol.* 2007;99:453-461.
105. Demoly P, Emminger W, Rehm D, Backer V, Tommerup L, Kleine-Tebbe J. Effective treatment of house dust mite-induced allergic rhinitis with 2 doses of the SQ HDM SLIT-tablet: results from a randomized double-blind, placebo-controlled phase III trial. *J Allergy Clin Immunol.* 2015;137:444-451.
106. de Bot CMA, Moed H, Berger MY, et al. Sublingual immunotherapy not effective in house dust mite-allergic children in primary care. *Pediatr Allergy Immunol.* 2012;23:151-159.
107. Didier A, Malling HJ, Worm M, et al. Optimal dose, efficacy, and safety of once-daily sublingual immunotherapy with a 5-grass pollen tablet for seasonal allergic rhinitis. *J Allergy Clin Immunol.* 2007;120:1338-1345.
108. Didier A, Melac M, Montagut A, Lheritier-Barrand M, Tabar A, Worm M. Agreement of efficacy assessments for five-grass pollen sublingual tablet immunotherapy. *Allergy.* 2009;64:166-171.
109. Didier A, Malling HJ, Worm M, et al. Post-treatment efficacy of discontinuous treatment with 300IR 5-grass pollen sublingual tablet in adults with grass pollen-induced allergic rhinoconjunctivitis. *Clin Exp Allergy.* 2013;43:568-577.
110. Durham SR, Yang WH, Pedersen MR, Johansen N, Rak S. Sublingual immunotherapy with once-daily grass allergen tablets: a randomized controlled trial in seasonal allergic rhinoconjunctivitis. *J Allergy Clin Immunol.* 2005;117:802-809.
111. Durham SR, Riis B. Grass allergen tablet immunotherapy relieves individual seasonal eye and nasal symptoms, including nasal blockage. *Allergy.* 2007;62:954-957.
112. Durham SR, Emminger W, Kapp A, et al. Long-term clinical efficacy in grass pollen-induced rhinoconjunctivitis after treatment with SQ-standardized grass allergy immunotherapy tablet. *J Allergy Clin Immunol.* 2009;125:131-138.
113. Durham SR, G. T. Investigators. Sustained effects of grass pollen AIT. *Allergy.* 2011;66(Suppl 95):50-52.
114. Durham SR, Emminger W, Kapp A, et al. SQ-standardized sublingual grass immunotherapy: confirmation of disease modification 2 years after 3 years of treatment in a randomized trial. *J Allergy Clin Immunol.* 2012;129:717-725.
115. Drachenberg KJ, Pfeiffer P, Urban E. Sublingual immunotherapy – Results from a multi-centre, randomised, double-blind, placebo-controlled study with a standardised birch and grass/rye pollen extract. [German]. *Allergologie.* 2001;24:525-534.
116. Feliziani V, Lattuada G, Parmiani S, Dall'Aglio PP. Safety and efficacy of sublingual rush immunotherapy with grass allergen extracts. A double blind study. *Allergy.* 1992;47:281-290.
117. Frolund L, Durham SR, Calderon M, et al. Sustained effect of SQ-standardized grass allergy immunotherapy tablet on rhinoconjunctivitis quality of life. *Allergy.* 2010;65:753-757.
118. Guez S, Vatrinet C, Fadel R, Andre C. House-dust-mite sublingual-swallow immunotherapy (SLIT) in perennial rhinitis: a double-blind, placebo-controlled study. *Allergy.* 2000;55:369-375.
119. Halcken S, Agertoft L, Seidenberg J, et al. Five-grass pollen 300IR SLIT tablets: efficacy and safety in children and adolescents. *Pediatr Allergy Immunol.* 2010;21:970-976.
120. Horak F, Stübner P, Berger UE, Marks B, Toth J, Jäger S. Immunotherapy with sublingual birch pollen extract. A short-term double-blind placebo study. *J Investig Allergol Clin Immunol.* 1998;8:165-171.
121. Horak F, Ziegelmayer P, Ziegelmayer R, et al. Early onset of action of a 5-grass-pollen 300-IR sublingual immunotherapy tablet evaluated in an allergen challenge chamber. *J Allergy Clin Immunol.* 2009;124:471-477.e1.
122. Hirsch T, Sahn M, Leupold W. Double-blind placebo-controlled study of sublingual immunotherapy with house dust mite extract (D.pt.) in children. *Paediatr Allergy and Immunol.* 1997;8:21-27.
123. Hordijk GJ, Antvelink JB, Luwema RA. Sublingual immunotherapy with a standardised grass pollen extract: a double-blind, placebo-controlled study. *Allergol Immunopathol (Madr).* 1998;26:234240.
124. Ibañez MD, Kaiser F, Knecht R, et al. Safety of specific sublingual immunotherapy with SQ standardized grass allergen tablets in children. *Pediatr Allergy Immunol.* 2007;18:516-522.
125. Ippoliti F, De Santis W, Volterrani A, et al. Immunomodulation during sublingual therapy in allergic children. *Pediatr Allergy Immunol.* 2003;14:216-221.
126. Kaluzinska-Parzyszek I, Majak P, Jerzynska J, Smejda K, Stelmach I. Sublingual immunotherapy is effective and safe in children. *Allergy Asthma Immunol.* 2011;16:139-144.



127. La Rosa M, Ranno C, Andre C, Carat F, Tosca MA, Canonica GW. Double-blind placebo-controlled evaluation of sublingual-swallow immunotherapy with standardized *Parietaria judaica* extract in children with allergic rhinoconjunctivitis. *J Allergy Clin Immunol*. 1999;104:425-432.
128. Marcucci F, Sensi L, Frati F, et al. Effects on inflammation parameters of a double-blind, placebo controlled one-year course of SLIT in children monosensitized to mites. *Allergy*. 2003;58:657-662.
129. Moreno-Ancillo A, Moreno C, Ojeda P, et al. Efficacy and quality of life with once-daily sublingual immunotherapy with grasses plus olive pollen extract without uposing. *J Investig Allergol Clin Immunol*. 2007;17:399-405.
130. Mosbech H, Canonica GW, Backer V, et al. SQ house dust mite sublingually administered immunotherapy tablet (ALK) improves allergic rhinitis in patients with house dust mite allergic asthma and rhinitis symptoms. *Ann Allergy Asthma Immunol*. 2014;114:134-140.
131. Mosges R, Bruning H, Hessler HJ, Gotz G, Knaussmann HG. Sublingual immunotherapy in pollen-induced seasonal rhinitis and conjunctivitis: a randomized controlled trial. *Acta Dermatovenerol Alp Panon Adriat*. 2007;16:143-148.
132. Okubo K, Gotoh M, Fujieda S, et al. A randomized double-blind comparative study of sublingual immunotherapy for cedar pollinosis. *Allergol Int*. 2008;57:265-275.
133. Ott H, Sieber J, Brehler R, et al. Efficacy of grass pollen sublingual immunotherapy for three consecutive seasons and after cessation of treatment: the ECRIT study.[Erratum for *Allergy*. 2009;64:179-86]. *Allergy*. 2009;64:1394-1401.
134. Nelson HS, Oppenheimer J, Vatsia GA, Buchmeier A. A double-blind, placebo-controlled evaluation of sublingual immunotherapy with standardized cat extract. *J Allergy Clin Immunol*. 1993;92:229-236.
135. Pajno GB, Vita D, Parmiani S, Caminiti L, La Grutta S, Barberio G. Impact of sublingual immunotherapy on seasonal asthma and skin reactivity in children allergic to *Parietaria* pollen treated with inhaled fluticasone propionate. *Clin Exp Allergy*. 2003;33:1641-1647.
136. Palma-Carlos AG, Santos AS, Branco-Ferreira M, et al. Clinical efficacy and safety of preseasonal sublingual immunotherapy with grass pollen carbamylated allergoid in rhinitic patients. A double-blind, placebo-controlled study. *Allergol Immunopathol (Madr)*. 2006;34:194-198.
137. Panzner P, Petrás M, Sýkora T, Lesná I. Double-blind, placebo-controlled evaluation of grass pollen specific immunotherapy with oral drops administered sublingually or supralingually. *Respir Med*. 2008;102:1296-1304.
138. Passalacqua G, Albano M, Fregonese L, et al. Randomised controlled trial of local allergoid immunotherapy on allergic inflammation in mite-induced rhinoconjunctivitis. *Lancet*. 1998;351:629-632.
139. Passalacqua G, Albano M, Riccio A, et al. Clinical and immunologic effects of a rush sublingual immunotherapy to *Parietaria* species: a double-blind, placebo-controlled trial. *J Allergy Clin Immunol*. 1999;104:964-968.
140. Passalacqua G, Pasquali M, Ariano R, et al. Randomized double-blind controlled study with sublingual carbamylated allergoid immunotherapy in mild rhinitis due to mites. *Allergy*. 2006;61:849-854.
141. Pfaar O, Klimek L. Efficacy and safety of specific immunotherapy with a high-dose sublingual grass pollen preparation: a double-blind, placebo-controlled trial. *Ann Allergy Asthma Immunol*. 2008;100:256-263.
142. Pradalier A, Basset D, Claudel A, et al. Sublingual-swallow immunotherapy (SLIT) with a standardized five-grass-pollen extract (drops and sublingual tablets) versus placebo in seasonal rhinitis. *Allergy*. 1999;54:819-828.
143. Purello-D'Ambrosio F, Gangemi S, Isola S, et al. Sublingual immunotherapy: a double-blind, placebo-controlled trial with *Parietaria judaica* extract standardized in mass units in patients with rhinoconjunctivitis, asthma, or both. *Allergy*. 1999;54:968-973.
144. Queiros MG, Silva DA, Siman IL, et al. Modulation of mucosal/systemic antibody response after sublingual immunotherapy in mite-allergic children. *Pediatr Allergy Immunol*. 2013;24:752-761.
145. Rak S, Yang WH, Pedersen MR, Durham SR. Once-daily sublingual allergen-specific immunotherapy improves quality of life in patients with grass pollen-induced allergic rhinoconjunctivitis: a double-blind, randomised study. *Qual Life Res*. 2006;16:191-201.
146. Rolinck-Werninghaus C, Wolf H, Liebke C, Baars JC, Lange J, Kopp MV. A prospective, randomized, double-blind, placebo-controlled multi-centre study on the efficacy and safety of sublingual immunotherapy (SLIT) in children with seasonal allergic rhinoconjunctivitis to grass pollen. *Allergy*. 2004;59:1285-1293.
147. Sabbah A, Hassoun S, Le Sellin J, Andre C, Sicard H. A double-blind, placebo-controlled trial by the sublingual route of immunotherapy with a standardized grass pollen extract. *Allergy*. 1994;49:309-313.
148. Stelmach I, Kaluzinska-Parzyszek I, Jerzynska J, Stelmach P, Stelmach W, Majak P. Comparative effect of pre-seasonal and continuous grass sublingual immunotherapy in children. *Allergy*. 2012;67:312-320.
149. Roder E, Berger MY, Groot H, Wijk RG. Immunotherapy in children and adolescents with allergic rhinoconjunctivitis: a systematic review (Structured abstract). *Pediatr Allergy Immunol*. 2008;19:197-207.
150. Tari MG, Mancino M, Monti G. Efficacy of sublingual immunotherapy in patients with rhinitis and asthma due to house dust mite. A double-blind study. *Allergol Immunopathol (Madr)*. 1990;18:277-284.
151. Valovirta E, Jacobsen L, Ljørring C, Koivikko A, Savolainen J. Clinical efficacy and safety of sublingual immunotherapy with tree pollen extract in children. *Allergy*. 2006;61:1177-1183.
152. Van Niekerk CH, De Wet JI. Efficacy of grass-maize pollen oral immunotherapy in patients with seasonal hay-fever: a double-blind study. *Clin Allergy*. 1987;17:507-513.
153. Vourdas D, Syrigou E, Potamianou P, et al. Double-blind, placebo-controlled evaluation of sublingual immunotherapy with standardized olive pollen extract in pediatric patients with allergic rhinoconjunctivitis and mild asthma due to olive pollen sensitization. *Allergy*. 1998;53:662-672.
154. Wang DH, Chen L, Cheng L, et al. Fast onset of action of sublingual immunotherapy in house dust mite-induced allergic rhinitis: a multicenter, randomized, double-blind, placebo-controlled trial. *Laryngoscope*. 2013;123:1334-1340.
155. Wahn U, Klimek L, Ploszczuk A, et al. High-dose sublingual immunotherapy with single-dose aqueous grass pollen extract in children is effective and safe: a double-blind, placebo-controlled study. *J Allergy Clin Immunol*. 2012;130:886-893.
156. Wahn U, Tabar A, Kuna P, et al. Efficacy and safety of 5-grass-pollen sublingual immunotherapy tablets in pediatric allergic rhinoconjunctivitis. *J Allergy Clin Immunol*. 2009;123:160-166.
157. Hylander T, Larsson O, Petersson-Westin U, Eriksson M, Kumlien Georén S, Wingqvist O. Intralymphatic immunotherapy of pollen-induced rhinoconjunctivitis: a double-blind placebo-controlled trial. *Respir Res*. 2016;17:10:1-9.
158. Senti G, Cramer R, Kuster D, Johansen P, Martinez-Gomez J, Graf N. Intralymphatic immunotherapy for cat allergy induces tolerance after only 3 injections. *J Allergy Clin Immunol*. 2012;129:1290-1296.

159. Didier A, Malling HJ, Worm M, Horak F, Sussman GL. Prolonged efficacy of the 300IR 5-grass pollen tablet up to 2 years after treatment cessation, as measured by a recommended daily combined score. *Clin Transl Allergy*. 2015;5:12.
160. Meadows A, Kaambwa B, Novielli N, et al. A systematic review and economic evaluation of subcutaneous and sublingual allergen immunotherapy in adults and children with seasonal allergic rhinitis. *Health Technol Assess*. 2013;17:27.
161. Canonica GW, Poulsen PB, Vestenbaek U. Cost-effectiveness of GRAZAX for prevention of grass pollen induced rhinoconjunctivitis in Southern Europe. *Respir Med*. 2007;101:1885-1894.
162. Keiding H, Jorgensen KP. A cost-effectiveness analysis of immunotherapy with SQ allergen extract for patients with seasonal allergic rhinoconjunctivitis in selected European countries. *Curr Med Res Opin*. 2007;23:1113-1120.
163. Bachert C, Vestenbaek U, Christensen J, Griffiths UK, Poulsen PB. Cost-effectiveness of grass allergen tablet (GRAZAX) for the prevention of seasonal grass pollen induced rhinoconjunctivitis – a Northern European perspective. *Clin Exp Allergy*. 2007;37:772-779.
164. Poulsen PB, Pedersen KM, Christensen J, Vestenbaek U. Economic evaluation of a tablet-based vaccination against hay fever in Denmark. *Ugeskr Laeger*. 2008;170:138-142.
165. Dranitsaris G, Ellis AK. Sublingual or subcutaneous immunotherapy for seasonal allergic rhinitis: an indirect analysis of efficacy, safety and cost. *J Eval Clin Pract*. 2014;20:225-238.
166. Ronaldson S, Taylor M, Bech PG, Shenton R, Bufe A. Economic evaluation of SQ-standardized grass allergy immunotherapy tablet (Grazax) in children. *Clin Outcomes Res*. 2014;6:187-196.
167. Westerhout KY, Verheggen BG, Schreder CH, Augustin M. Cost effectiveness analysis of immunotherapy in patients with grass pollen allergic rhinoconjunctivitis in Germany. *J Med Econ*. 2012;15:906-917.
168. Verheggen B, Westerhout K, Schreder C, Augustin M. Health economic comparison of SLIT allergen and SCIT allergoid immunotherapy in patients with seasonal grass-allergic rhinoconjunctivitis in Germany. *Clin Transl Allergy*. 2015;5:1.
169. Reinhold T, Brüggjenjürgen B. Cost-effectiveness of grass pollen SCIT compared with SLIT and symptomatic treatment. *Allergo J Int*. 2017;26:7-15.
170. Ruggeri M, Oradei M, Frati F, et al. Economic evaluation of 5-grass pollen tablets versus placebo in the treatment of allergic rhinitis in adults. *Clin Drug Investig*. 2013;33:343-349.
171. Berto P, Passalacqua G, Crimi N, et al. Economic evaluation of sublingual immunotherapy vs symptomatic treatment in adults with pollen-induced respiratory allergy: the Sublingual Immunotherapy Pollen Allergy Italy (SPAI) study. *Ann Allergy Asthma Immunol*. 2006;97:615-621.
172. Brüggjenjürgen B, Reinhold T, Brehler R, et al. Cost-effectiveness of specific subcutaneous immunotherapy in patients with allergic rhinitis and allergic asthma. *Ann Allergy Asthma Immunol*. 2008;101:316-324.
173. Schadlich PK, Brecht JG. Economic evaluation of specific immunotherapy versus symptomatic treatment of allergic rhinitis in Germany. *Pharmacoeconomics*. 2000;17:37-52.
174. Omnes LF, Bousquet J, Scheinmann P, et al. Pharmacoeconomic assessment of specific immunotherapy versus current symptomatic treatment for allergic rhinitis and asthma in France. *Eur Ann Allergy Clin Immunol*. 2007;39:148-156.
175. Peterson K, Gyrd-Hansen D, Dahl R. Health-economic analyses of subcutaneous specific immunotherapy for grass pollen and mite allergy. *Allergol Immunopathol (Madr)*. 2005;33:296-302.
176. Pokladnikova J, Krcmova I, Vlcek J. Economic evaluation of sublingual vs subcutaneous allergen immunotherapy. *Ann Allergy Asthma Immunol*. 2008;100:482-489.
177. Nasser S, Vestenbaek U, Beriot-Mathiot A, Poulsen PB. Cost-effectiveness of specific immunotherapy with Grazax in allergic rhinitis co-existing with asthma. *Allergy*. 2008;63:1624-1629.
178. Ariano R, Berto P, Incorvaia C, Di Cara G, Boccardo R, La Grutta S. Economic evaluation of sublingual immunotherapy vs symptomatic treatment in allergic asthma. *Ann Allergy Asthma Immunol*. 2009;103:254-259.
179. Almagro E, Asensio O, Bartolome J, Bosque M, De La Hoz B, Dolz i. Multicenter drug surveillance of sublingual immunotherapy in allergic patients. *Allergol Immunopathol (Madr)*. 1994;23:153-159.
180. Brehler R, Klimek L, Pfaar O, Hauswald B, Worm M, Bieber T. Safety of a rush immunotherapy build-up schedule with depigmented polymerized allergen extracts. *Allergy Asthma*. 2010;31:31-38.
181. Cardona R, Lopez E, Beltrán J, Sánchez J. Safety of immunotherapy in patients with rhinitis, asthma or atopic dermatitis using an ultra-rush buildup. A retrospective study. *Allergol Immunopathol (Madr)*. 2014;42:90-95.
182. Casanovas M, Martin R, Jiménez C, Caballero R, Fernández-Caldas E. Safety of an ultra-rush immunotherapy build-up schedule with therapeutic vaccines containing depigmented and polymerized allergen extracts. *Int Arch Allergy Immunol*. 2006;139:153-158.
183. Casanovas M, Martin R, Jiménez C, Caballero R, Fernández-Caldas E. Safety of immunotherapy with therapeutic vaccines containing depigmented and polymerized allergen extracts. *Clin Exp Allergy*. 2007;37:434-440.
184. Pfaar O, Mösges R, Hörmann K, Klimek L. Safety aspects of Cluster immunotherapy with semi-depot allergen extracts in seasonal allergic rhinoconjunctivitis. *Eur Arch Otorhinolaryngol*. 2010;267:245-250.
185. Yi H, Liu Y, Ye J, Yu J. Clinical observation of the adverse effects of standardized dust mite allergen preparation in the treatment of allergic rhinitis. *J Clin Otorhinolaryngol Head Neck Surg*. 2014;28:1870-1872.
186. Cohen J. *Statistical Power Analysis for the Behavioral Sciences*. Vol. 2nd ed. Hillsdale, NJ: Erlbaum; 1988.
187. Pfaar O, Demoly P, Gerth van Wijk R, et al. Recommendations for the standardization of clinical outcomes used in allergen immunotherapy trials for allergic rhinoconjunctivitis: an EAACI Position Paper. *Allergy*. 2014;69:854-867.
188. Pfaar O, Bastl K, Berger U, et al. Defining pollen exposure times for clinical trials of allergen immunotherapy for pollen-induced rhinoconjunctivitis – an EAACI position paper. *Allergy*. 2017;72: 713-722.

## SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

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